



# *Ficus* spp. (Figs) and their anticancer potential: A systematic review of laboratory studies and traditional uses in the Philippines



Kathleen Laum Cabanlit<sup>1,2\*</sup>, Mark Anthony Jariol Torres<sup>1,2,3</sup>, Cesar Guinanao Demayo<sup>1,2,3</sup>

<sup>1</sup>Department of Biological Sciences, Mindanao State University-Iligan Institute of Technology, Iligan City, Philippines

<sup>2</sup>Center of Integrative Health, Premier Research Institute of Science and Mathematics, MSU- Iligan Institute of Technology, Iligan City, Philippines

<sup>3</sup>School of Interdisciplinary Studies/Institute of Peace and Development in Mindanao, MSU-Iligan Institute of Technology, Iligan City, Philippines

## ARTICLE INFO

**Article Type:**  
Review

### Article History:

Received: 8 Dec. 2024

Revised: 22 Jan. 2025

Accepted: 27 Jan. 2025

published: 1 Apr. 2025

### Keywords:

*Ficus* species

Anticancer

Cytotoxicity

Ethnomedicine

Traditional medicine

Plant-derived compound

## ABSTRACT

**Introduction:** *Ficus* species (figs) have been traditionally utilized for managing various diseases, including cancer, a leading cause of global morbidity and mortality. Despite their widespread use, it remains an underexplored genus in modern pharmacognosy. This review documents species used as anticancer remedies in the Philippines and synthesizes global laboratory evidence supporting their anticancer properties. This approach bridges traditional knowledge with modern research, offering new insights into the therapeutic potential of the genus.

**Methods:** A systematic review was conducted following PRISMA guidelines. Relevant studies were retrieved from electronic databases (ScienceDirect, PubMed) and expanded search (Google Scholar). Database filters and keywords, such as “anticancer” and “*Ficus*”, were optimized with Boolean operators and wildcards to refine search results. Study quality was evaluated using the Mixed Methods Appraisal Tool (MMAT).

**Results:** A total of 76 articles met the inclusion criteria, with 58 classified as high quality and 18 as moderate-high quality. Five key species, namely *Ficus septica*, *Ficus elastica*, *Ficus congesta*, *Ficus concinna*, and *Ficus botryocarpa*, were recorded for local traditional use, predominantly prepared as leaf decoctions. Laboratory evidence identified bioactive compounds such as terpenoids, flavonoids, alkaloids, phenolics, and species-specific compounds, exhibiting cytotoxic effects against multiple cancer types through mechanisms such as cell cycle arrest, apoptosis, antioxidant modulation, and metastasis inhibition.

**Conclusion:** This review highlights the potential of *Ficus* spp. as a source of anticancer agents, aligning traditional uses with laboratory evidence. Findings provide a foundation for further exploration of *Ficus*-based therapies in clinical applications.

### Implication for health policy/practice/research/medical education:

This review provides information on the potential of *Ficus* species as anticancer agents, encouraging further research into their clinical use. Policymakers should support funding for research and collaborations between scientists and traditional healers to bridge ethnomedicine and modern medicine. In medical practice, *Ficus*-based medicines may serve as complementary cancer treatments, especially in resource-limited settings.

**Please cite this paper as:** Cabanlit KL, Torres MAJ, Demayo CG. *Ficus* spp. (Figs) and their anticancer potential: A systematic review of laboratory studies and traditional uses in the Philippines. J Herbmed Pharmacol. 2025;14(2):133-152. doi: 10.34172/jhp.2025.52854.

## Introduction

Cancer remains a leading cause of morbidity and mortality worldwide, with its prevalence steadily increasing in both developed and developing countries (1). Current estimates of approximately 20 million new cases annually are projected to double by 2070, driven by demographic

shifts and rising cancer incidence across regions (2). This growing global burden accentuates the urgent need for effective prevention, treatment, and therapeutic strategies to combat this complex disease. However, traditional cancer treatments such as chemotherapy, radiation, and surgery, are often associated with significant side

\*Corresponding author: Kathleen Laum Cabanlit,  
Email: [kathleen.cabanlit@g.msuiit.edu.ph](mailto:kathleen.cabanlit@g.msuiit.edu.ph)

effects and limitations, prompting a growing interest in alternative and complementary therapies, particularly those derived from natural sources.

Throughout history, medicinal plants have been vital natural resources in improving health outcomes, forming the foundation of herbal therapy. As a biodiversity hotspot, the Philippines boasts a rich tradition of herbal medicine, making its local ethnobotanical knowledge particularly significant (3). These longstanding practices provide valuable ethnobotanical information that can guide scientific research. Medicinal claims rooted in tradition are often based on generations of empirical observation, offering a practical starting point for identifying plants with bioactive compounds and therapeutic potential. Investigating these claims presents an opportunity to uncover novel therapeutic agents and mechanisms, advancing the scientific understanding of natural products and their latent applications in modern medicine.

*Ficus* species, commonly known as figs, have nutritional value and diverse medicinal properties (4). *Ficus* species hold a unique place in Filipino culture, where their use is deeply intertwined with spiritual beliefs and healing rituals. They are often regarded as sacred, believing that they aid in healing. This belief is integral to the practices of traditional healers or shamans who emphasize the connection between spirituality and physical well-being (5,6).

Despite their rich traditional usage and cultural significance, *Ficus* species remain underexplored compared to other medicinal plants. Systematic evaluations of their anticancer potential are limited, which is likely due to the genus's vast biodiversity, fragmented ethnobotanical knowledge, and the chemical complexity of its secondary metabolites (7). Furthermore, research has historically prioritized plants with well-established pharmacological profiles, leaving *Ficus* spp. as an underutilized resource for drug discovery.

This study aimed to address this gap by systematically reviewing laboratory studies on the anticancer properties of *Ficus* spp., providing an inventory of the potential anticancer compounds identified in different parts of the plants, their mechanisms of action, and the types of cancer for which these compounds exhibit anticancer-related effects. The emphasis on plant parts is due to the understanding that different parts may contain distinct compounds, similar to traditional practices that use various parts of the plants. Additionally, the review provides an overview of the *Ficus* species used in the Philippines as traditional remedies for cancer and the methods employed to harness their perceived medicinal potential. Unlike previous reviews (8), this study provides a novel perspective by integrating global laboratory studies with local ethnobotanical knowledge from the Philippines, highlighting the therapeutic potential of *Ficus* spp. through a combined traditional and scientific lens (ethnobotany and herbal medicine pharmacology).

Through this dual approach, patterns and correlations between traditional and scientific information can be deduced, unveiling the pharmacological activities of *Ficus* spp. while situating their anticancer properties within the broader context of traditional medicine. This perspective supports drug discovery efforts and validates and preserves traditional knowledge systems, which are at risk due to modernization and habitat destruction. Moreover, the review of available studies allows for noting research gaps, such as variability in experimental methods and challenges in translating findings into clinical applications. Ultimately, this comprehensive evaluation seeks to strengthen the foundation for future research, conservation efforts, and drug development pipelines, reinforcing *Ficus* species as promising candidates for novel anticancer therapies.

## Review Methodology

### Review team

One reviewer (KC), initially screened the titles and abstracts for relevance. Articles deemed potentially suitable were retrieved in full text and assessed independently by CD and MAT, with KC performing a subsequent re-assessment. In cases where there was disagreement on article inclusion, a consensus was made through a thorough and collaborative review by all evaluators. The Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) were used to guide the process of record identification, screening, and inclusion (Figure 1).

### Data sources and design

The electronic databases ScienceDirect and PubMed were systematically searched, with additional expanded searches, including forward and backward citations, conducted via Google Scholar. These databases were selected for their extensive coverage and strong reputation for reliability, which are commonly favored by other authors conducting literature reviews. Additionally, they were chosen for accessibility, given that other academic databases require institutional subscriptions that the authors' institute did not have. The systematic conduct of this review was referred from the PRISMA design (9) from inception to November 15, 2024.

### Search strategy plan and optimization process

An iterative trial-and-error approach was used to refine search strategies and achieve ideal results. The titles and abstracts of five relevant studies were initially searched to identify potential search keywords. These keywords were added to the 'Advanced Search' fields in the databases and expanded the search. Relevant filters were applied to the fields such as titles, abstracts, and full texts. Wildcards were adjusted where necessary to improve search flexibility and accommodate different terminologies. Box 1 shows the final set of optimized keywords for these sources.

The eligibility criteria established were as follows:

## Box 1. Optimized keywords

## PubMed

(((Ficus) AND (cancer)) NOT (nanoparticle)) NOT (review)

## NCBI filter

Text availability: Free full text

## ScienceDirect

"Ficus", "anticancer" -review -nanoparticle

## SD Filter

Article type: Research articles

## Google Scholar

"Ficus", "anticancer" -review -nanoparticle

written in English; tested on a single *Ficus* species only (no herbal combinations); the genus of plant/s tested was/were *Ficus*; natural compounds from *Ficus* spp.; analysis focused on plant extract itself, not symbionts; and directly tested for anticancer activity. These criteria were initially established upon initial screening of articles for inclusion. However, in the course of extracting data, other reasons for exclusion were distinguished and presented in the PRISMA flow diagram (Figure 1).

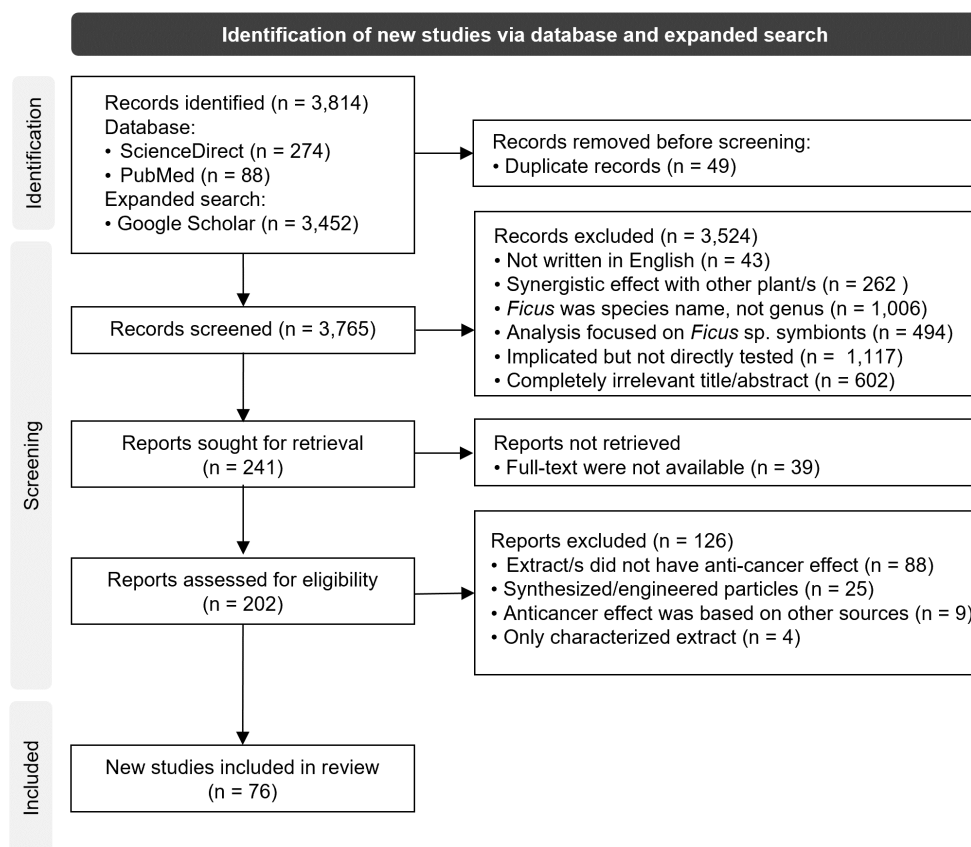
The traditional anticancer use of *Ficus* spp. in the Philippines was taken from our previous results (10). Meanwhile, the group/class of compounds were mentioned

in each article where they were cited. In a few cases where the compound class was not mentioned, other articles were referred for proper identification. Microsoft Excel was the spreadsheet tool used to tabulate, organize, and sort all the data, consequently transferred to be presented in tabular form using Microsoft Word. The R package and Shiny application (11) were used to generate the PRISMA flow diagram.

The initial data extraction from databases and search engines yielded 3,814 records, with 274 articles from ScienceDirect, 88 from PubMed, and 3452 from manual searches—including forward and backward citations—via Google Scholar. After removing duplicates, 3765 articles remained for further screening. Of these, 3524 were excluded for reasons outlined in Figure 1, leaving 241 articles for retrieval. Full-text availability allowed 202 articles to be assessed for eligibility, with 76 ultimately included in the review.

## Quality assessment

The quality of the included studies was assessed independently by the reviewers using the Mixed Methods Appraisal Tool (MMAT) (12), specifically under the “quantitative non-randomized studies” category. This categorization was appropriate for this review as they typically report results such as IC<sub>50</sub> values. However, to avoid bias, these values were not currently considered since



**Figure 1.** PRISMA flow diagram of the review on the ethnobotany and anticancer potential of *Ficus* spp.

different *Ficus* species and cancer cell lines were tested. Hence, quantitative comparisons were unwarranted. In using MMAT, each criterion was equivalent to a score of 1 for yes, 0 for no, and CT for can't tell. In cases where confusion arised, a thorough discussion was made between the reviewers to reach a consensus. After consolidating input from the three reviewers, 58 articles (76.32%) were classified as high quality ( $\geq 0.80$ ), while 18 articles (23.68%) fell into the moderate-high quality range (0.66–0.79), as evaluated using MMAT ver. 2018.

## Results

### Taxonomy

Modern taxonomy places *Ficus* spp. within the following classifications: Kingdom Plantae (all plants), Subkingdom Viridiplantae (green plants), Phylum Tracheophyta (vascular plants), Subphylum Spermatophytina (seed plants), Class Magnoliopsida (dicotyledons), Order Rosales (plants like roses, figs, and mulberries), Family Moraceae (mulberries), and Genus *Ficus* (figs) (13).

### Botany

The genus *Ficus*, belonging to the Moraceae family, comprises more than 750 species classified into four subgenera and 20 sections (14), including well-known varieties such as the common fig (*Ficus carica*) and the rubber tree (*Ficus elastica*). These trees and shrubs are characterized by their large, often glossy leaves, which can be lobed or unlobed depending on the species. Their leaves produce a milky latex when damaged, which has traditional medicinal and adhesive properties (15,16).

Many *Ficus* species also develop aerial roots, which, in species like the banyan tree (*Ficus benghalensis*), form massive and complex structures that provide additional support.

*Ficus* encompasses diverse growth forms, including trees, shrubs, epiphytes, and vines, easily recognized by their unique reproductive structure, the syconium. This enclosed inflorescence houses numerous small male and female flowers that, upon pollination, develop into multiple fruits known as figs (17,18). Each *Ficus* species is typically pollinated by a single species of wasp from the family Agaonidae (19), illustrating a mutualistic coevolutionary process. These fig wasps pollinate the flowers while simultaneously laying eggs inside the syconium, facilitating seed production for the *Ficus* and providing a protected environment for their larvae (20).

**Traditional anticancer uses of *Ficus* spp. in the Philippines**  
Previous ethnobotanical studies have documented traditional uses of *Ficus* as an anticancer agent in the Philippines (Table 1).

While there was no direct counterpart to the biomedical concept of 'cancer' in local terminology, traditional healers and knowledge systems recognized it through distinct signs and symptoms. These include (1) bukol or 'lump,' which refers to any abnormal growth or swelling in the body, (2) sakit sa laman or 'disease of the flesh,' describing chronic and worsening pain in the body, and (3) pagpayat or 'wasting,' which signifies unexplained weight loss and general physical decline. These indicators help local healers identify potential cancer cases through observable

**Table 1.** Traditional use of *Ficus* species as potential anticancer agents in the Philippines based on local practices

<i>Ficus</i> spp.	Common name	Local name	Plant part used	Mode of preparation	Administration	Claimants	Sources
<i>botryocarpa</i>	Cluster fig	Busyong	Bark	Decoction	Boil 10 inches long and 5 inches wide bark in an ample amount of water; drink 1/2 glass often.	Subanen tribe (Lapuyan, ZDS)	(21)
<i>concinna</i>	Elegant fig	Pulang balite	Leaves; bark; roots	Decoction	Drink the decoction once a day in thrice a week for 2 months (5-7 glasses)	Manobo tribe (Agusan del Sur)	(22)
<i>congesta</i>	Red-leaf fig	Kalfagang	Leaves	Poultice	Apply to the affected part	B'laan tribe (Mt. Matutum)	(23)
<i>elastica</i>	Rubber fig	Dakit	Leaves	Decoction	Drink the decoction	Locals of Catmon, Toledo, and Alcoy, Cebu	(24)
<i>elastica</i>	Rubber fig	Balite na dako; Goma	Leaves	Decoction	Drink the decoction	Pagadian City	(25)
<i>elastica</i>	Rubber fig	Balite na dako; Goma	Leaves	Poultice	Poultice	Tribes of the Zamboanga peninsula (Visayan)	(26)
<i>septica</i>	Hauili fig tree	Hawili	Leaves	Poultice	Apply as salve	Kanawan Aytas	(27)
<i>septica</i>	Hauili fig tree	Marabutan	Leaves; bark	Decoction	Drink the decoction	Subanen Tribe (Sindangan)	(28)
<i>septica</i>	Hauili fig tree	Hawili	Leaves; bark	Decoction	Drink the decoction	Maranao (Bubong, LDS)	(29)
<i>septica</i>	Hauili fig tree	Ikmo/Buyo	Leaves; bark	Decoction	Drink the decoction	Iligan City	(30)
<i>septica</i>	Hauili fig tree	Marabutan	Leaves; bark	Decoction	Drink the decoction	Higaonon (Rogongon, Iligan City)	(31)
<i>septica</i>	Hauili fig tree	Marabutan	Leaves	Decoction	Drink the decoction	Dinagat Islands	(32)

physical signs, relying on a symptom-based understanding rather than a specific term equivalent to 'cancer'.

In contrast to more common ethnobotanical uses for ailments like coughs, wounds, or headaches, these cancer-related symptoms are relatively few, resulting in fewer documented plants for anticancer use. Ethnobotanical surveys in the Philippines identified five *Ficus* species specifically noted for their anticancer uses: *Ficus septica* (Hauili), *Ficus elastica* (Rubber fig), *Ficus congesta* (Red-leaf fig), *Ficus concinna* (Elegant fig), and *Ficus botryocarpa*

(Cluster fig), listed according to their frequency of occurrence.

#### Laboratory studies on phytochemical constituents of *Ficus* spp. with documented anticancer activity

Previous studies have identified several bioactive compounds in *Ficus* species with documented anticancer activity (Table 2). Reviewing these compounds is essential for identifying potential therapeutic agents, validating traditional medicinal practices, and understanding the

**Table 2.** Documented anticancer phytochemicals in *Ficus* spp.

Plant part	Compound class	<i>Ficus</i> spp.	Compound	References
Alkaloid		<i>septica</i>	(+)-Isotylocrebrine; Tylocrebrine; (+)-tylophorine; Ficuseptine; 14R-hydroxyisocrebrine N-oxide; Tylophorine; Indole	(33-35)
		<i>fistulosa</i>	Fistulopsene B; Tengechlorenine; Tengerensine	(36,37)
		<i>hispida</i>	Hispilosine	(38)
Alkane		<i>crocata</i>	Hexatriacontane	(39)
Carboxylic acid		<i>carica</i>	Benzoic acid	(40)
Carotenoid		<i>virens</i>	Lycopersen	(41)
Coumarin		<i>carica</i>	Bergapten; Psoralen	(42)
Ester		<i>virens</i>	Dibutyl phthalate	(41)
		<i>ovata</i>	Hexadecanoic acid	(43)
Fatty acid		<i>crocata</i>	10,13,13-trimethyl-11-tetradecen-1-ol acetate	(44)
		<i>carica</i>	Apigenin; Luteolin, Lutin, Quercetin	(40)
		<i>deltoidea</i>	Isovitexin; Vitexin	(45)
Flavonoid		<i>pseudopalma</i>	Quercetin	(46)
		<i>microcarpa</i>	Gingerglycolipid B; (2S)-2,3-O-dioctadeca-9Z,12Z,15Z-trienoylglyceryl-O-β-D-galactopyranoside; (2S)-2,3-O-dioctadeca-9Z,12Z-dienoylglyceryl-O-β-D-galactopyranoside; (2S)-3-O-octadeca-9Z,12Z,15Z-trienoylglyceryl-6'-O-(α-D-galactopyranosyl)-β-D-galactopyranoside; (2S)-2,3-O-dioctadeca-9Z,12Z,15Z-trienoylglyceryl-6'-O-(α-D-galactopyranosyl)-β-D-galactopyranoside; (2S)-2,3-O-dioctadeca-9Z,12Z-dienoylglyceryl-6'-O-(α-D-galactopyranosyl)-β-D-galactopyranoside	(47)
Leaves	Ketone	<i>religiosa</i>	Benzophenone	(41)
		<i>elasticoides</i>	6,10,14-trimethyl-2-pentadecanone	(43)
		<i>ovata</i>	6,10,14-trimethyl-2-pentadecanone	(43)
		<i>rumphii</i>	3-(2-hydroxyphenyl)-1-(piperidin-1-yl) propan-1-one	(48)
		<i>crocata</i>	Vitamin E	(39)
	Phenolic	<i>benghalensis</i>	Carvacrol	(41)
		<i>religiosa</i>	Tocopherol	(41)
		<i>rumphii</i>	1-isopentyl-3,4-dioxomethylene-2-phenol	(48)
		<i>auriculata</i>	2,4-ditertiary-butyl-phenol	(49)
		<i>carica</i>	Protocatechuic acid; Gallic acid	(40,50)
Sterol	<i>crocata</i>	β-sitosterol	(39,44)	
	<i>carica</i>	β-sitosterol	(40,51)	
	<i>ingens</i>	β-sitosterol	(51)	
	<i>hispida</i>	β-sitosterol	(52)	
	<i>benghalensis</i>	Phytol	(41)	
Terpenoid		<i>elasticoides</i>	(E)-phytol; β-carophyllene	(43)
		<i>ovata</i>	(E)-phytol; Carophyllene oxide	(43)
		<i>natalensis</i>	(E)-phytol	(43)
		<i>crocata</i>	Lup-20(29)-en-3-ol acetate (3β); Phytol; Squalene; Lup-20(29)-en-3-ol-acetate; Lupeol	(39, 44)
		<i>pseudopalma</i>	Lupeol	(46)
	<i>microcarpa</i>	2(S)-3-O-octadeca-9Z,12Z,15Z-trienoylglyceryl-O-β-D-galactopyranoside	(47)	
	<i>hispida</i>	Lup-20(29)-en-3-ol-acetate	(52)	
	<i>pomifera</i>	3β-Acetoxyurs-3-ene; 3β-hydroxyurs-3-ene	(53)	
	<i>deltoidea</i>	Ursolic acid	(54)	



Table 2. Continued

Plant part	Compound class	<i>Ficus</i> spp.	Compound	References	
Fruit	Amide	<i>carica</i>	(2-benzhydrylsulfonyl)-N-hydroxyacetamide)-Na	(55)	
		<i>hispida</i>	3'-formyl-5,7-dihydroxy-4'-methoxyisoflavone; Alpinumisoflavone; Isowigtheone hydrate; $\beta$ -sitosterol 3-O- $\beta$ -D-glucopyranoside	(56)	
		<i>bubu</i>	Biochanin A	(57)	
	Flavonoid	<i>carica</i>	Cyanidin 3-O-glucoside; Quercetin; Quercetin 3- $\beta$ -glucoside; Rutin	(58)	
		<i>altissima</i>	Isoflavone 2; Isoflavone 3; Isolupalbigenin; Isowigtheone; Lupinalbin; Luteolin	(59)	
		<i>vasta</i>	Quercetin	(60)	
	Phenolic	<i>sycomorus</i>	Quercetin-3-rutinoside	(61)	
		<i>hispida</i>	Gallic acid; Chlorogenic acid; Chlorogenic acid methyl ester	(56)	
		<i>carica</i>	Ellagic acid; Ferulic acid; Trans-cinnamic acid	(58)	
	Terpenoid	<i>vasta</i>	Gallic acid	(60)	
		<i>palmata</i>	Caffeic acid; Catechin; Coumaric acid; Gallic acid	(62)	
		<i>bubu</i>	Trans-resveratrol; Piceid	(57)	
	Roots	Alkaloid	<i>hispida</i>	Betulinic acid	(56)
			<i>carica</i>	Ficutirucin A; Ficutirucin B; Ficutirucin C; Ficutirucin F; Ficutirucin G; Ficutirucin I	(63)
		Alkaloid	<i>septica</i>	10S,13aR-tylocrebrine N-oxide; 13aR-isotylocrebrine; 13aR-tylocrebrine; 13aR-tylophorine; Ficuseptine A	(64,65)
<i>elastica</i>			Elasticamide	(65)	
Flavonoid		<i>elastica</i>	Ficososide B; Ficososide B (peracetylated); Biochanin A	(65,66)	
		<i>beeheyana</i>	Epicatechin; Rutin	(64,66)	
Lignin derivative		<i>hirta</i>	Ficuslignin B	(67)	
Phenolic		<i>beeheyana</i>	Caffeic acid; Chlorogenic acid; Gallic acid; p-coumaric acid; p-hydroxybenzoic acid; Protocatechuic acid	(66)	
		<i>elastica</i>	Elastiquinone	(65)	
Terpenoid		<i>microcarpa</i>	Plectranthoic acid	(68)	
Stem	Alkaloid	<i>septica</i>	10S,13aR-isotylocrebrine N-oxide; 10S,13aR-tylocrebrine N-oxide; Tylophorine	(69)	
		<i>foveolata</i>	Foveolide A; Foveolide B	(70)	
Bark	Alkaloid	<i>fistulosa</i>	Fistulopsine A	(35)	
		<i>drupacea</i>	5-O-methylatifolin	(71)	
	Flavonoid	<i>glumosa</i>	6-prenylapigenin; Luteolin	(72)	
		<i>religiosa</i>	Phenolic acid	(73)	
	Saponin	<i>glumosa</i>	Dongnoside E	(72)	
	Terpenoid	<i>drupacea</i>	Epifriedelanol; Epilupeol acetate; Friedelin; Oleanolic acid	(71)	
		<i>glumosa</i>	$\beta$ -amyryne; Lanosta-7,24-dien-3-one; Lupeol	(72)	
Latex	Flavonoid	<i>exasperata</i>	Ursolic acid	(74)	
		<i>carica</i>	Quercetin	(75,76)	
	Phenolic	<i>carica</i>	Caffeic acid; Ferulic acid; p-coumaric acid; Protocatechuic acid	(75,76)	
<i>dubia</i>		Chlorogenic acid; Quinic acid; Syringoylquinic acid	(77)		

mechanisms through which these compounds exert their effects on cancer cells. This exploration can lead to the discovery of novel drug leads and highlight the ecological and cultural significance of *Ficus* species. Additionally, understanding their phytochemical diversity can inspire the development of synergistic therapies that enhance the efficacy of conventional treatments while potentially reducing side effects.

The leaves contained a wide variety of compounds, with

terpenoids being the most abundant (18 occurrences), followed by alkaloids (11), flavonoids (7), and phenolics (7) (Figure 2). Other compounds belonging to classes of glycosides (6), ketones (5), sterols (5), coumarins (2), fatty acids (2), alkane (1), carboxylic acid (1), carotenoid (1), and ester (1) were also reported.

The most common compounds in fruits were flavonoids (14 occurrences), mainly from *Ficus altissima* and *Ficus carica*, alongside phenolics (11) and terpenoids (7). In the



first isolated from *Ficus altissima* and *Ficus hispida*, respectively. Isowighteone significantly inhibited nitric oxide (NO) production compared to the nonsteroidal anti-inflammatory drug (NSAID) indomethacin (59). Other compounds isolated in the same study included isolupalbigenin, lupinalbin, luteolin, and two unidentified isoflavones, all of which demonstrated antiproliferative and cytotoxic effects against breast (MDA-MB-231) and liver (HepG2) cancer cell lines. Isowighteone hydrate, on the other hand, exhibited cytotoxic activity against the human promyelocytic leukemia (HL-60) cell line by activating caspases 3, 8, and 9 (56).

*Ficus elastica* and *Ficus beecheyana* contained flavonoids in their roots, with a new compound named Ficoside B being isolated for the first time from the aerial roots of *Ficus elastica* (65). Flavonoids were also found in the bark of *Ficus drupacea* and *Ficus glumosa*; in the leaves of *Ficus deltoidea*, *Ficus pseudopalma*, and *Ficus carica*, as well as in the latex of *Ficus carica*. Notably, quercetin was found in nearly all parts of *Ficus carica* studied (40,58,75,76).

Phenolic compounds were found in the fruits, latex, roots, leaves, and bark of *Ficus* spp. The leaves had the most diverse species containing potent phenolic compounds including *Ficus carica*, *Ficus rumphii*, *Ficus religiosa*, *Ficus crocata*, *Ficus benghalensis*, and *Ficus auriculata* (39-41,48-50). Four species (*Ficus palmata*, *Ficus hispida*, *Ficus carica*, and *Ficus vasta*) had phenolics in their fruits, with gallic acid being common to all of them (56,60,62). *Ficus carica* and *Ficus dubia* contained phenolics in their latex (75-77), while *Ficus religiosa* had phenolic acid in its bark (73). *Ficus beecheyana* roots also contained phenolic compounds including caffeic acid, gallic acid, and protocatechuic acid (66).

Alkaloids were most prevalent in the leaves, with *Ficus septica* having most of the compounds, including newly isolated phenanthroindolizidine alkaloids, such as Ficuseptines B-D, (+)-isotylocrebrine, tylocrebrine, and (+)-tylophorine. In addition to the leaves, alkaloids were also found in the roots and stems of *Ficus septica*. The leaves of *Ficus fistulosa* also contained a species-specific alkaloid, Fistulopsene B, while its bark contained a similar alkaloid, Fistulopsene A. Both compounds belong to a unique class of septicine-type alkaloids, which have been shown to arrest cancer cells in the G1 phase without inducing apoptosis (35). Moreover, the leaves of *Ficus hispida* also contained a species-specific alkaloid, Hispiloscine (38).

Terpenoids were identified in the least diverse plant parts, limited to the leaves, fruits, and bark. Most of the species, including *Ficus crocata*, *Ficus pomifera*, *Ficus microcarpa*, *Ficus deltoidea*, *Ficus pseudopalma*, *Ficus ovata*, *Ficus natalensis*, *Ficus elasticoides*, and *Ficus benghalensis*, contain terpenoids in their leaves, with phytol and lupeol being common among them. *Ficus carica* and *Ficus hispida* had terpenoids in their fruits, with the unique isolation of Ficutirucins, a type of tirucallane

triterpenoid, from *Ficus carica* which exhibited cytotoxic activity against several cancer cell lines. Additionally, *Ficus drupacea*, *Ficus glumosa*, and *Ficus exasperata* contain terpenoids in their bark, including oleanolic acid and epilupeol acetate, both of which exhibit antiproliferative activity (71). Other terpenoids from *Ficus drupacea*, such as epifriedelanol and friedelin, have also been reported to exhibit high cytotoxicity against the tested cancer cell lines (71).

#### *Ficus* spp. anticancer phytochemical profile and mechanisms of action

The anticancer activity of the bioactive compounds has been mechanistically varied, depending on the compounds present, the pathways involved, and the type of cancer cell line targeted. The diversity in mechanisms allows these compounds to act on multiple cellular processes, thereby contributing to their potential as anticancer agents (Table 3).

#### Comparative phytochemical profiles and therapeutic potential of *Ficus* spp.

Various *Ficus* species exhibit significant anticancer activities on various cancer cell lines, with distinct pharmacodynamic contributions from different plant parts. Notably, *Ficus carica* has notable activities. For instance, its latex extracts induced apoptosis and enhanced temozolomide (TMZ) efficacy in glioblastoma cells by upregulating Let-7D expression (75). Meanwhile, its leaf extracts downregulated GATA3 in breast cancer cells (MDA-MB-231), promoting cell cycle arrest while exhibiting antiproliferative, anti-metastatic, and antiangiogenic properties by modulating ERK2, CREB, and AKT2 pathways (84). Furthermore, *Ficus carica* fruits demonstrated complex apoptosis-inducing effects in ovarian (SKOV-3) and breast (AMJ-13) cancer cells through the upregulation of P53, Caspase-8, and Bax (55).

*Ficus altissima*, rich in flavonoids, demonstrated antiproliferative activity and nitric oxide production inhibition on breast (MDA-MB-231) and liver (HepG2) cancer cells (59), emphasizing its potential for targeting cancer cell growth. *Ficus auriculata* showed cytotoxic effects on gastric (AGS) cancer cells and induced cell cycle changes and apoptosis in lung (A549) cancer cells through a caspase-independent pathway, attributed to its phenolic content (49,78). Similarly, species like *Ficus deltoidea* and *Ficus drupacea*, containing flavonoids and terpenoids, demonstrated significant cytotoxic and apoptotic effects (45,54,71,90).

Antiproliferative effects were also exhibited by *Ficus benghalensis*, which was abundant in carotenoids, esters, ketones, phenolics, and terpenoids, against breast (MDA-MB-468) and colon (Colo320) cancer cells (41,80). It also demonstrated neuroprotective activity against oxidative stress in neuroblastoma cells (81, 82). Notably, novel compounds with anticancer activity were isolated



**Table 3.** Phytochemical profiles of different parts of *Ficus* species and their anticancer pharmacodynamics

<i>Ficus</i> spp.	Plant part	Plant preparation	Compound/ Extract	Mechanism of action	Ref.
<i>altissima</i>	Fruit	Air-dried	Flavonoids	Antiproliferative activity and NO production inhibition on breast (MDA-MB-231) and liver (HepG2) cancer cells	(59)
<i>auriculata</i>	Leaves	Shade-dried	Phenolics	Cytotoxic activity on gastric (AGS) cancer cells	(49)
<i>auriculata</i>	Fruits	Freeze-dried	Extract	Induction of cell cycle profile changes and apoptosis induction via caspase-independent pathway on lung (A549) cancer cells	(78)
<i>awkeotsang</i>	Seeds	Fresh	Extract	Induction of cell cycle arrest at G2/M phase and apoptosis induction via mitochondrial pathway involved with exceeding ROS level	(79)
<i>beecheയാ</i>	Roots	Air-dried	Flavonoids, phenolics	Growth inhibition of leukemia (HL-60) cells	(66)
<i>beecheയാ</i>	Roots	Air-dried	Flavonoids, phenolics	Induction of apoptosis via <i>Fas</i> - and mitochondrial-mediated pathway and reduction of mitochondrial membrane potential of leukemia (HL-60) cells	(66)
<i>benghalensis</i>	Leaves	Dried	Carotenoid, ester, ketone, phenolics, terpenoids	Antiproliferative activity on breast (MDA-MB-468) cancer cells	(41)
<i>benghalensis</i>	Root	Shade-dried	Extract	Antiproliferative activity on colon (Colo320) cancer cells	(80)
<i>benghalensis</i>	Bark	Air-dried	Extract	Neuroprotective activity (against oxidative environment generated by H <sub>2</sub> O <sub>2</sub> treatment) on neuroblastoma (SK N SH) cells	(81)
<i>benjamina</i>	Extract	Dried	Coumarins	High binding affinity of compounds to PARP-1 (an anticancer therapy target)	(82)
<i>bubu</i>	Fruit	Air-dried	Flavonoids, stilbenes, terpenoids	Peracetylation-induced increase in antiproliferative activity and cytotoxic activity on lung (A549) and skin (SKMEL-28) cancer cells	(57)
<i>carica</i>	Latex	Fresh	Flavonoids, phenolics	Cell proliferation inhibition (reduced invasion via induction of Let-7D expression), apoptosis induction, enhancement of TMZ effect, and inhibition of angiogenesis on glioblastoma cells	(75)
<i>carica</i>	Leaves	Fresh	Coumarins	Antiproliferative activity, downregulation of breast cancer marker gene GATA3, cell cycle arrest, and decrease in cell mobility on breast (MDA-MB-231) cancer cells	(42)
<i>carica</i>	Fruit	Air-dried	Amide	Increased ROS synthesis, reduction of MMP via <i>Bax</i> upregulation by P53, Caspase-8, and Caspase-9 and Bcl-2 downregulation, inhibition of proliferation, and induction of apoptosis via activated P53 and caspase-8 signaling on ovary (SKOV-3) and breast (AMJ-13) cancer cells	(55)
<i>carica</i>	Fruit	Air-dried	Flavonoids, phenolics	Antiproliferative activity by alleviating plasma cytokine levels and phenolic composition on breast (MDA-468) cancer cells	(58)
<i>carica</i>	Extract	Dried	Phenolic	Cytotoxic activity (decreased cell viability) and induction of apoptosis on bone (MG-63) and colon cancer (HT-29) cancer cells	(50)
<i>carica</i>	Leaves	Shade-dried	Sterols	Cytotoxic activity on liver (HepG2), kidney (HEK-293), and breast (MCF-7; MDA-MB-231) cancer cells	(51)
<i>carica</i>	Fruit	Air-dried	Terpenoids	Cytotoxic activity on liver (HepG2), breast (MCF-7) and bone (U2OS) cancer cells	(63)
<i>carica</i>	Leaves	Air-dried	Flavonoids	The regulatory substance of MCM protein expression in anticancer mechanisms	(83)
<i>carica</i>	Latex	Fresh	Flavonoids, phenolics	Apoptosis induction, invasion prevention via Let-7d expression in glioblastoma (e U-138 MG, T98G, and U-87 MG) cells	(76)
<i>carica</i>	Extract	Extract	Flavonoids, terpenoids	High binding affinity of compounds to PARP-1 (an anticancer therapy target)	(82)
<i>carica</i>	Leaves	Air-dried	Carboxylic acid, flavonoids, phenolics, sterols	Cytotoxic activity, anti-colonization activity, cell cycle arrest at S phase, increase in ROS leading to loss of mitochondrial membrane potential, downregulation of TP53, Bcl-2, CDK1, 5, and 9, on liver (HepG2) cancer cells	(40)
<i>carica</i>	Latex (leaves)	Fresh	Extract	Antiproliferative activity, anti-metastatic, genotoxic, and cytotoxic activity, and downregulation of ERK2, CREB, and AKT2 on breast (MDA-MB-231) cancer cells	(84)

Table 3. Continued

<i>Ficus</i> spp.	Plant part	Plant preparation	Compound/ Extract	Mechanism of action	Ref.
<i>carica</i>	Peel, pulp, leaves, fruit, latex	Air-dried	Extract	Inhibition of proliferation of colorectal (HT-29) and colon (HCT-116) cancer cells; apoptosis induction on colorectal cancer cells	(85)
<i>carica</i>	Fruit, leaves, latex	Air-dried	Extract	Decreased cell viability of cervical (HeLa) cancer cells	(86)
<i>carica</i>	Leaves	Sun-dried	Extract	Anti-angiogenesis activity is implicated via the number of blood vessels and VEGF expression of CAM of embryonated chicken eggs	(87)
<i>carica</i>	Fruits	Air-dried	Extract	Cytotoxic activity, apoptosis induction, DNA damage, Increase in iCa <sup>2+</sup> levels on breast (MCF-7) cancer cells	(88)
<i>carica</i>	Leaves	Air-dried	Extract	Apoptosis induction via proteolytic cleavage of PARP, downregulation of Bcl-2, and upregulation of <i>Bax</i> , cell viability reduction, colony formation inhibition via caspase- and mitochondrial-dependent apoptotic pathways on oral (FaDu) cancer cells	(89)
<i>crocata</i>	Leaves	Fresh	Alkane, phenolics, sterols, terpenoids	Cytotoxic activity, reduction of cancer cell migration, reduction of invasion capacity and secretion of MMP-2 and MMP-9, reduction of actin polymerization, and probably EMT on the breast (MDA-MB-231; MCF-7) cancer cells	(39)
<i>crocata</i>	Leaves	Air-dried	Fatty acids, sterols, terpenoids	Antiproliferative activity, apoptosis induction, and increased p53, procaspase-8, and procaspase-3 expression in breast (MDA-MB-231) cancer cells	(44)
<i>deltoidea</i>	Leaves	Fresh	Flavonoids	Cell reduction of prostate (DU145) cancer cells by apoptosis	(45)
<i>deltoidea</i>	Leaves	Fresh	Flavonoids	Decreased cell viability on prostate (DU145) cancer cell lines	(45)
<i>deltoidea</i>	Leaves	Oven-dried	Terpenoids	Cytotoxic activity on colorectal (HCT-116) and breast (MDA-MB-231) cancer cells	(54)
<i>deltoidea</i>	Leaves	Oven-dried	Terpenoids	Inhibition of micro-vessels outgrowth in rat aortic rings and inhibition of chick CAM vascularization	(54)
<i>deltoidea</i>	Leaves	Oven-dried	Terpenoids	Cytotoxic activity against colon (HCT-116) and breast (MDA-MB-231) cancer cells	(54)
<i>deltoidea</i>	Leaves	Air-dried	Extract	Antiproliferative activity and apoptosis induction via upregulation of Fas1, Bax, Cdk-1, TNF- $\alpha$ and Cdk-2 and downregulation of Bcl-2 and Tp53 on breast (MCF-7, MDA-MB-231, HCC 1937) and colon (HCT-116) cancer cells	(90)
<i>deltoidea</i>	Leaves	Oven-dried	Extract	Apoptosis induction, promotion of cell detachment, and inhibition of proliferation through DNA fragmentation on ovarian (A2780) cancer cells	(91)
<i>deltoidea</i>	Leaves	Air-dried	Extract	Regression on tumor progression, reduced incidence of oral squamous cell carcinoma (OSCC), decreased expression of cyclin D1 (tumor marker), and increased expression of $\beta$ -catenin and e-cadherin antibodies reduced the expression of the TWIST1 and RAC1 (EMT) and downregulated the COX-2 and EGFR (angiogenesis, metastasis, and chemoresistance)	(92)
<i>drupacea</i>	Bark	Air-dried	Flavonoids, terpenoids	Antiproliferative activity on cervical (HeLa), breast (MCF-7), leukemia (Jurkat), and colorectal (HT-29, T24) cancer cells	(71)
<i>dubia</i>	Latex	Fresh	Phenolics	Antiproliferative activity, cell cycle arrest by NF- $\kappa$ B, cyclin D1, CDK4, p21 downregulation, selective apoptosis induction by down-regulating NF- $\kappa$ B and Bcl-xl and up-regulating Bid, Bak, cleaved caspase-7 and caspase-3 in colorectal (HCT-116 and HT-29) cancer cells.	(77)
<i>elastica</i>	Liana	Air-dried	Alkaloids, saponins, tannins, terpenoids	Antiproliferative activity on oligodendroglioma (Hs683) and breast (MCF-7) cancer cells	(65)
<i>elastica</i>	Roots	Oven-dried	Amide, flavonoids, quinone	Antiproliferative activity on brain (U373, Hs683), breast (MCF-7), lung (A549), and skin (B16F10, SK-MEL-28) cancer cells	(65)
<i>elasticoidea</i>	Leaves	Air-dried	Ketone, terpenoids	Compounds associated with anticancer mechanisms via apoptotic pathways	(43)
<i>exasperata</i>	Bark	Air-dried	Terpenoids	Cytotoxic activity on cervical (KB-3-1) and colon (HT-29) cancer cells	(74)

Table 3. Continued

<i>Ficus</i> spp.	Plant part	Plant preparation	Compound/ Extract	Mechanism of action	Ref.
<i>fistulosa</i>	Bark, leaves	Fresh	Alkaloids	Growth inhibition and cell cycle arrest in the G1 phase without apoptosis induction of colon (HCT-116) and breast (MCF-7) cancer cells	(35)
<i>fistulosa</i>	Leaves	Air-dried	Alkaloids	Cytotoxic activity on breast (MDA-MB-468, MDA-MB-231 and MCF-7) cancer cells	(38)
<i>foveolata</i>	Stems	Air-dried	Terpenoids	Cytotoxic activity on colorectal (SW620), liver (HepG2). Breast (BT474) and stomach (KATO-III) cancer cells	(70)
<i>glomerata</i>	Whole plant	Oven-dried	Extract	Antiproliferative activity on the liver (HepG2) and breast (MCF-7) cancer cells	(93)
<i>glumosa</i>	Bark	Air-dried	Flavonoids, saponins, terpenoids	Cytotoxic activity on the prostate (PC-3) and fibrosarcoma (HT1080) cancer cells	(72)
<i>heterophylla</i>	Roots	Shade-dried	Alkaloids, flavonoids, phenolics, tannins	Compounds implicated in anticancer mechanisms via apoptotic pathways	(94)
<i>hirta</i>	Roots	Air-dried	Lignin derivative	Cytotoxic activity and apoptosis induction via JNK/p38 MAPK pathway on liver (HepG2) cancer cells	(67)
<i>hirta</i>	Roots	Air-dried	Extract	Decreased cell viability and apoptosis induction via cell cycle arrest at sub-G1 phase on cervical (HeLa) cancer cells	(95)
<i>hispidia</i>	Leaves	Shade-dried	Fatty alcohol, sterols, terpenoids	Compounds implicated in anticancer mechanisms via apoptotic pathways	(52)
<i>hispidia</i>	Extract	Dried	Flavonoids, terpenoids	High binding affinity of compounds to PARP-1 (an anticancer therapy target)	(82)
<i>hispidia</i>	Fruits	Air-dried	Flavonoids, glycoside, phenolics, sterols	Cancer chemopreventive activity, cytotoxic activity on leukemia (HL60), lung (A549), breast (SK-BR-3), oral (KB), cervical (HeLa), colon (HT-29), and liver (Hep-G2) cancer cells, and apoptosis induction via both the mitochondrial and death receptor-mediated pathways by activation of caspases-3, -8, and -9 in leukemia (HL60) cells	(56)
<i>hispidia</i>	Bark, leaves	Fresh	Alkaloids	Antiproliferative activity on breast (MDA-MB-231, MCF-7), lung (A549), and colon (HCT-116) cancer cells	(36)
<i>hispidia</i>	Leaves	Air-dried	Extract	Cytotoxic activity via apoptosis induction and cell cycle arrest at G0/G1 phase on colorectal (HT-29) cancer cells	(96)
<i>ingens</i>	Leaves	Shade-dried	Sterols	Cytotoxic activity on the liver (HepG2) and breast (MDA-MB-231) cancer cells	(51)
<i>krishnae</i>	Bark	Dried	Extract	Inhibition of proliferation and colony formation of breast (MCF-7) cancer cells	(97)
<i>lacor</i>	Extract	Dried	Flavonoids, terpenoids	Contains compound/s with high binding affinity to PARP-1 (an anticancer therapy target)	(82)
<i>microcarpa</i>	Roots	Air-dried	Terpenoids	Antiproliferative activity by G0/G1 phase cell cycle arrest attributed to cyclin kinase inhibitors upregulation and apoptosis induction in AMPK-dependent manner on prostate (DU145, CW22Rv1, PC3) nerve (NB26), and skin (A375) cancer cells.	(98)
<i>microcarpa</i>	Leaves	Air-dried	Glycosides	Inhibition of TNF- $\alpha$ induced IL-8 secretion in the colon (HT-29) cancer cells.	(47)
<i>microcarpa</i>	Roots	Air-dried	Terpenoids	EMT inhibition leads to decreased migration and reverse effect, epithelial markers induction and decreased mesenchymal markers, inhibition of Rac1/NEDD9 pathway on prostate (DU145, PC3, NA22, and NB26) cancer cells	(68)
<i>natalensis</i>	Leaves	Air-dried	Ketone, terpenoids	Compounds associated with anticancer mechanisms via apoptotic pathways	(43)
<i>ovata</i>	Leaves	Air-dried	Fatty acid, ketone, terpenoids	Compounds associated with anticancer mechanisms via apoptotic pathways	(43)
<i>palmata</i>	Fruits	Fresh	Phenolics	Cytotoxic activity on cervical (C33A) cancer cells	(62)
<i>pandurata</i>	Aerial parts	Fresh	Polysaccharide	Antiproliferative activity and apoptosis induction via upregulation of caspase-3 and cleaved-PARP on cervical (HeLa) cancer cells	(99)

Table 3. Continued

<i>Ficus</i> spp.	Plant part	Plant preparation	Compound/ Extract	Mechanism of action	Ref.
<i>pomifera</i>	Leaves	Air-dried	Terpenoids	Cytotoxic activity on lung (A549) cancer cells	(53)
<i>pseudopalma</i>	Leaves	Air-dried	Extract	Inhibition of cell growth and apoptosis induction on liver (HepG2) cancer cells	(100)
<i>pseudopalma</i>	Leaves	Air-dried	Flavonoids, terpenoids	Cytotoxic activity decreased cell viability and apoptosis induction in prostate (PRST2) cancer cells	(46)
<i>pumila</i>	Leaves	Air-dried	Extract	Decrease in number and size of tumorspheres, induction of apoptosis, cell cycle arrest at G2/M phase, upregulation of p53, p21, and GADD45A genes, downregulation of cyclin B1 and cyclin D1 genes on liver (HepG2) cancer cells	(101)
<i>religiosa</i>	Bark	Shade-dried	Phenolics	Cytotoxic activity on cervical (HeLa, SiHa) cancer cells	(73)
<i>religiosa</i>	Extract	Dried	Sterols	Chemo-preventive properties that promote antioxidant gene activity through regulation of Nrf2 factor by competing for receptor site with Keap1 during stressed conditions	(102)
<i>religiosa</i>	Leaves	Air-dried	Alkaloids, flavonoids, phenolics, sterols, tannins	G1 phase cell cycle arrest, sub-g0 phase induction, apoptosis induction, decrease in mitochondrial membrane potential, elevation of Caspase 9 activity, cleavage of prominent Caspases and PARP, intracellular ROS generation, photosensitizing activity through rapid mitochondrial transmembrane potential loss and partial Caspase activation, growth inhibition associated with Bax translocation and mitochondria-mediated apoptosis with the activation of Caspase 9-dependent cascade on breast (MCF-7) cancer cells.	(103)
<i>religiosa</i>	Leaves	-	Carotenoid, ester, ketone, phenolics, terpenoids	Antiproliferative activity on breast (MDA-MB-468) cancer cells	(41)
<i>religiosa</i>	Leaves	Air-dried	Extract	Antiproliferative activity on breast (MCF-7) cancer cells	(104)
<i>religiosa</i>	Leaves	Shade-dried	Extract	Antiproliferative activity on leukemia (Jurkat) cells	(105)
<i>rumphii</i>	Leaves	Air-dried	Ketone, phenolics	Antiproliferative activity on lung (A549), liver (HepG2), and breast (MDA-MB-361) cancer cells	(48)
<i>salicifolia</i>	Latex (leaves)	Fresh	Extract	Antiproliferative activity, anti-metastatic, genotoxic, and cytotoxic activity, and upregulation of ERK2, CREB, and AKT2 at the transcriptional level on breast (MDA-MB-231) cancer cells	(84)
<i>septica</i>	Leaves	Air-dried	Alkaloids	Cytotoxic activity on stomach and nasopharyngeal (NUGC and HONE-1) cancer cells	(33)
<i>septica</i>	Leaves	Air-dried	Alkaloids	Cytotoxic activity on cervical (HeLa) cancer cells	(34)
<i>septica</i>	Stem	Air-dried	Alkaloids	Cytotoxic activity on stomach and nasopharyngeal (NUGC and HONE-1) cancer cells	(69)
<i>septica</i>	Roots	Air-dried	Alkaloids	Cytotoxic activity on stomach and nasopharyngeal (NUGC and HONE-1) cancer cells	(64)
<i>septica</i>	Leaves	Air-dried	Alkaloids	Cytotoxic activity on breast (T47D) cancer cells	(37)
<i>septica</i>	Leaves	Fresh	Extract	Decrease cell viability on breast (T47D) cancer cell	(106)
<i>septica</i>	Leaves	Air-dried	Extract	Apoptosis induction via p53-independent pathway	(107)
<i>septica</i>	Leaves	Air-dried	Extract	Cytotoxic activity on lung (A549) and colon (HCT-116) cancer cells	(108)
<i>sycomorus</i>	Fruit	Air-dried	Flavonoids	Compounds implicated in anticancer activities	(61)
<i>vasta</i>	Fruit	Air-dried	Flavonoids, Phenolics	Reduction of cell viability, disruption of the cell membrane, leaking of cell debris, and apoptotic bodies development in lung and breast (A549; MDA-MB-231) cancer cells	(60)
<i>virens</i>	Leaves	Dried	Carotenoid, Ester, Ketone, Phenolics, Terpenoids	Antiproliferative activity on breast (MDA-MB-468) cancer cells	(41)

from *Ficus foveolata* stems, such as the eudesmane-type sesquiterpene eoveolide A and the sesquiterpenoid dimer Foveolide B. Foveolide A showed moderate cytotoxicity against colorectal (SW620), liver (HepG2), breast (BT474), and gastric (KATO-III) cancer cell lines, while Foveolide B exhibited cytotoxicity specifically towards colorectal cancer cells (70).

Some species have been shown to have both direct actions on cancer cells and indirect effects, such as enhancing chemotherapeutic efficacy. For instance, *Ficus carica* offers a broad range of therapeutic effects. It contains flavonoids, phenolics, and sterols with cell proliferation inhibition, apoptosis induction, and downregulation of cancer-related genes across various cancer types, such as glioblastoma, breast, and colon cancers (75-89). It is also recognized for its ability to enhance chemotherapy efficacy, such as potentiating TMZ effects on glioblastoma cells (75). Similarly, *Ficus religiosa* possesses a diverse array of compounds, including phenolics, sterols, alkaloids, and flavonoids, which are potent in inducing G1 phase arrest and apoptosis in breast cancer cells (41,103,104) and exhibiting chemo-preventive actions through antioxidant regulation (102). *Ficus microcarpa*, with its terpenoid and glycoside content, prevents the migration of cancer cells (metastasis) inhibiting cancer cell migration and metastasis by epithelial-mesenchymal transition (EMT) inhibition (47,68).

While each *Ficus* species possesses a unique phytochemical profile, they collectively present a diverse range of anticancer mechanisms, including apoptosis induction (95,96,98,101), inhibition of cancer metastasis (46,68,92), and regulation of key cancer-related pathways (78,79,89,107). The differences in their bioactive compounds and mechanisms underscore the potential for targeted cancer therapies using specific *Ficus* species, as well as the need for further research to explore their full therapeutic potential.

### Mechanisms of anticancer actions

Figure 2 summarizes the anticancer mechanisms of *Ficus* species extracts and compounds based on the studies included in this review. The extracts have exhibited their effects through apoptosis, cell cycle arrest, and tumor suppression. Apoptosis is induced via death receptor pathways and mitochondrial dysfunction, involving key regulators such as BAX, BAK, and caspases (78,89,90,103). The generation of reactive oxygen species (ROS) supplements this process, further inhibiting cancer cell proliferation (40,55,79).

Cell cycle arrest occurred through the downregulation of critical proteins like CDK1, CDK4, CDK9, and cyclin D1, which effectively halted cancer cell growth (90,92,98,101). Tumor progression and metastasis were also suppressed by reduced VEGF expression, which inhibited angiogenesis (87), and by decreased levels of MMP-2 and MMP-9, which limited cancer cell invasion

(39). Cytotoxicity was driven by mitochondrial damage, modulation of cancer-related genes (e.g., TP53, Bcl-2) (40,90), and chemopreventive effects through Nrf2-mediated antioxidant pathways (102). Collectively, these mechanisms represent an integrated response that inhibits tumor growth, metastasis, and cancer progression, highlighting the broad anticancer potential of *Ficus* species as demonstrated by the studies included in this review.

### Discussion

The Philippines' tropical climate provides year-round access to plant leaves, making them a sustainable resource for traditional healers (109). Leaves are preferred for their accessibility, ease of collection, minimal invasiveness, and richness in bioactive compounds. Decoction, a common traditional preparation, enhances the extraction of these compounds by utilizing their water-soluble nature, making medicinal properties more accessible (110-112). This method aids in breaking down cell walls through boiling, releasing active phytochemicals (113). Its simplicity makes decoction an essential practice in rural areas with limited healthcare access. In many Philippines' rural communities, figs are believed to house spirits and deities, highlighting their spiritual and cultural significance in healing rituals (7,8,114). Local healers often combine spiritual and physical approaches to health, making *Ficus* species central to holistic practices and alternative cancer treatments (115,116).

Plants, like all organisms, have natural defense systems to scavenge free radicals, but environmental changes lead to excessive free radical production, causing oxidative stress. This damage, in humans, can contribute to cancer (117,118). Bioactive compounds involved in plant defense against oxidative damage are typically alkaloids, phenolics, and terpenoids (119). These compounds, found in plants, play crucial roles in defense mechanisms and pharmaceutical research (120). Terpenoids, along with alkaloids and phenolics, are central to the chemical defense strategies of *Ficus* species (121). Alkaloids are toxic to herbivores and microorganisms, enhancing plant defense. This diversity of chemical defenses, crucial for addressing biotic and abiotic stresses, is also linked to their anticancer potential, targeting cellular DNA damage induced by carcinogens (122).

Flavonoids scavenge ROS and protect plants from environmental stressors like UV radiation and pathogens (123-126). In humans, they reduce oxidative stress, a key driver of DNA damage and cancer, while enhancing blood plasma antioxidant capacity and preventing LDL oxidation (127). *Ficus* fruits, rich in flavonoids, especially anthoxanthins and anthocyanins, play vital roles in seed protection and reproduction by attracting dispersers (128). The flavonoid content in *Ficus* fruits highlights their therapeutic potential, both in traditional medicine and as functional foods with health benefits.



The roots of *Ficus* species, though less diverse in compounds, are rich in alkaloids with high cytotoxic potential. This is due to structural features like a rigid phenanthrene structure and specific substitution patterns that enhance toxicity, supported by structure-activity relationship studies (129). The accumulation of alkaloids and terpenoids in the stem bark serves as chemical deterrents to herbivores and insects. Flavonoids and phenolics protect against UV radiation, oxidative damage, and fungal infections (130,131). However, the lower compound diversity in stems likely reflects their primary structural role rather than metabolite production.

*Ficus* spp. latex plays key roles due to its biochemical composition. Enzymes and metabolites in latex facilitate nutrient transport, ensuring efficient distribution within the plant (132). The phenolic and flavonoid content enhances the plant's protection, with phenolics providing antioxidant and antimicrobial properties and flavonoids offering anti-inflammatory and cytotoxic effects (133). Its sticky texture and irritants deter herbivores and pests. Some of these latex compounds also show anticancer activity (75,76), suggesting that *Ficus* latex not only supports the plant's survival but may also have therapeutic applications.

The traditional uses of *Ficus* species in the Philippines align with laboratory findings, further supporting their therapeutic potential. For example, *Ficus septica* and *Ficus elastica* decoctions and poultices, used for cancer symptoms such as external swelling (24–26), are supported by laboratory findings showing anti-inflammatory and antioxidant properties. These properties may help reduce oxidative stress and promote healing (65,66). The use of *Ficus botryocarpa* and *Ficus concinna* bark decoctions by the Subanen and Manobo tribes (21,22) was not supported by studies included in this review. However, extracts from other *Ficus* species' bark, such as *Ficus drupacea*, *Ficus exasperata*, *Ficus fistulosa*, *Ficus glumosa*, and *Ficus religiosa*, demonstrated significant cytotoxic effects against various types of cancer (35,71–74). This complex biochemical profile enables the bark to withstand external pressures, promoting the plant's resilience and enhancing its medicinal potential. The synthesis of these compounds is influenced by the bark's structural role, environmental exposure, and the need to protect vital tissues from damage, making it a reservoir of therapeutic molecules (134).

The wide range of compounds in *Ficus* species suggests that traditional preparation methods were developed to maximize the plant's therapeutic potential. Despite the lack of scientific knowledge about its chemical composition, locals utilized its benefits by applying various preparation techniques. This mirrors how modern extraction protocols in *in vitro* studies isolate potent compounds. Traditional methods like decoctions and salves correspond to the plant's bioactive compound variety. Decoction extract water-soluble compounds by

boiling leaves or bark, are ideal for treating systemic issues like internal inflammation. Salves, made by crushing plant materials for topical use, concentrate compounds found in the plant's sap or oils, providing localized relief for swelling. The alignment between traditional methods and the pharmacological effects of *Ficus* phytochemicals offers a scientific rationale for their use in managing conditions such as cancer, validating the effectiveness of these practices. These correlations emphasize how traditional knowledge complements scientific research, offering valuable insights into *Ficus* species' therapeutic uses. However, they also underline the need for further studies, particularly *in vivo* and clinical trials, to confirm the efficacy and pharmacokinetics of these preparations in modern medicine.

### Implications for therapeutic development

Variability in experimental methods, such as extraction techniques, solvents, and concentration, presents challenges in comparing results. The lack of *in vivo* studies limits understanding of pharmacokinetics and efficacy in living organisms. Translating *Ficus* findings into clinical applications faces additional hurdles, including the complexity of isolating active compounds, the need for standardization, and regulatory challenges. The diverse nature of *Ficus* metabolites may necessitate advanced formulation techniques to enhance bioavailability and ensure target-specific delivery.

Future research should prioritize clinical trials to evaluate the safety and efficacy of *Ficus* extracts, particularly for less-studied cancer types. Investigating molecular mechanisms, such as the modulation of signaling pathways involved in apoptosis and cell cycle regulation, will be critical to understanding their therapeutic potential. Moreover, studying underexplored *Ficus* species may lead to the discovery of novel bioactive compounds with distinct anticancer properties. This review reinforces the potential of *Ficus* species as valuable resources in cancer research and therapy, offering promising opportunities for future drug development.

### Conclusion

Five *Ficus* species—*Ficus septica*, *Ficus elastica*, *Ficus congesta*, *Ficus concinna*, and *Ficus botryocarpa*—have been traditionally used in the Philippines as cancer remedies, with leaf decoctions being the most common preparation. Laboratory studies confirm a diverse array of bioactive compounds, including terpenoids, alkaloids, flavonoids, and phenolics, distributed differently across plant parts. Terpenoids were most abundant in leaves, flavonoids in fruits, and alkaloids and phenolics in roots. Novel chemical compounds unique to specific species were also identified, highlighting the distinct phytochemical profiles of different plant parts that contribute to their medicinal properties.

A correlation between traditional knowledge and

scientific findings is evident, as laboratory studies support the therapeutic potential of *Ficus* species against cancer. These species demonstrated anticancer activity across a wide range of cell lines, including those from the breast, lung, liver, colon, prostate, cervix, ovary, stomach, kidney, oral cavity, blood, skin, bone, and brain. Their mechanisms of action include inducing cell cycle arrest, apoptosis, and anti-angiogenesis, modulating antioxidant genes, and inhibiting cancer growth and metastasis. However, research variability and the scarcity of *in vivo* studies present challenges in translating these findings into clinical applications. Key limitations include differences in experimental methods, difficulties isolating active compounds, standardizing preparation techniques, and enhancing bioavailability. Future research should prioritize clinical trials, further elucidation of molecular mechanisms, and exploration of understudied species to fully harness the anticancer potential of *Ficus*. By bridging traditional knowledge with contemporary scientific evidence, this study underscores the alignment between ethnobotanical practices and laboratory results, offering a robust foundation for advancing *Ficus* species as promising candidates for cancer research and therapy. *In vivo* studies remain crucial to validate these findings and unlock the full therapeutic potential of this underexplored genus.

#### Authors' contribution

**Conceptualization:** Cesar Guinanao Demayo and Mark Anthony Jariol Torres.

**Data curation:** Kathleen Laum Cabanlit, Cesar Guinanao Demayo, and Mark Anthony Jariol Torres.

**Formal analysis:** Kathleen Laum Cabanlit, Cesar Guinanao Demayo, and Mark Anthony Jariol Torres.

**Funding acquisition:** Kathleen Laum Cabanlit.

**Investigation:** Kathleen Laum Cabanlit, Cesar Guinanao Demayo, and Mark Anthony Jariol Torres.

**Methodology:** Kathleen Laum Cabanlit.

**Project administration:** Cesar Guinanao Demayo and Mark Anthony Jariol Torres.

**Resources:** Kathleen Laum Cabanlit.

**Software:** Kathleen Laum Cabanlit.

**Supervision:** Cesar Guinanao Demayo and Mark Anthony Jariol Torres.

**Validation:** Kathleen Laum Cabanlit, Cesar Guinanao Demayo, and Mark Anthony Jariol Torres.

**Visualization:** Kathleen Laum Cabanlit.

**Writing—original draft:** Kathleen Laum Cabanlit.

**Writing—review & editing:** Kathleen Laum Cabanlit and Cesar Guinanao Demayo.

#### Conflict of interests

The authors declare there is no conflict of interest.

#### Ethical considerations

The review followed the PRISMA guidelines, overseen

by the PRISMA Executive, to ensure transparency and methodological rigor. Ethical issues, including plagiarism, have been completely observed by the authors.

#### Funding/Support

The authors received funding from the Department of Science and Technology – Accelerated Science and Technology Human Resource Development Program (DOST-ASTHRDP) through Dr. Jayeel S. Cornelio, Director of DOST-SEI.

#### References

1. Lin L, Li Z, Yan L, Liu Y, Yang H, Li H. Global, regional, and national cancer incidence and death for 29 cancer groups in 2019 and trends analysis of the global cancer burden, 1990-2019. *J Hematol Oncol.* 2021;14(1):197. doi: 10.1186/s13045-021-01213-z.
2. Soerjomataram I, Bray F. Planning for tomorrow: global cancer incidence and the role of prevention 2020-2070. *Nat Rev Clin Oncol.* 2021;18(10):663-72. doi: 10.1038/s41571-021-00514-z.
3. Susaya-Garcia J, Borja N, Sevilla-Nastor J, Villanueva J, Peyraube N. An ethnobotanical study of medicinal plants and perceptions on plant biodiversity conservation in Leyte, Philippines. *J Hum Ecol.* 2018;7(1):26-42.
4. Alaman BB, Labajo-Villantes Y, Pito EC, Garrido AF, Villaneva GV, Talip OS, et al. New record of Philippine endemic *Ficus* species in Mt. Malindang, Mindanao, Philippines. *Int J Bot Stud.* 2020;5(4):193-6.
5. Celeste BL, Condino MP, Dadang RJ, Amoroso VB. Forest care, interconnectivity and maintenance of ecological resources among the Manobo-Matigsalug people of the Southern Philippines. *Environmental & Socio-economic Studies.* 2020;8(3):21-33. doi: 10.2478/environ-2020-0015.
6. Smith MC. *Gold and Wood: Material Culture and Ritual in Precolonial and Catholic Philippines.* Honolulu: University of Hawaii at Manoa; 2023. p. 120.
7. Lansky EP, Paavilainen HM. *Figs: The Genus Ficus.* CRC Press; 2010. p. 416.
8. Remadevi V, Mohan Lathika L, Sasikumar Sujatha A, Sreeharshan S. *Ficus* extract-a promising agent for antimammary tumorigenesis: a review on current status and future possibilities. *Phytother Res.* 2019;33(6):1597-603. doi: 10.1002/ptr.6348.
9. Page MJ, McKenzie JE, Bossuyt PM, Boutron I, Hoffmann TC, Mulrow CD, et al. The PRISMA 2020 statement: an updated guideline for reporting systematic reviews. *BMJ.* 2021;372:n71. doi: 10.1136/bmj.n71.
10. Cabanlit K, Torres MA, Demayo C. Traditional medicinal plants used as anti-cancer in the Philippines: a systematic ethnobotanical review. *Ethnobot Res Appl.* 2024;27:1-51.
11. Haddaway NR, Page MJ, Pritchard CC, McGuinness LA. PRISMA2020: an R package and Shiny app for producing PRISMA 2020-compliant flow diagrams, with interactivity for optimised digital transparency and Open Synthesis. *Campbell Syst Rev.* 2022;18(2):e1230. doi: 10.1002/cl2.1230.
12. Hong QN, Fàbregues S, Bartlett G, Boardman F, Cargo M, Dagenais P, et al. The Mixed Methods Appraisal Tool (MMAT) version 2018 for information professionals and researchers. *Educ Inf.* 2018;34(4):285-91. doi: 10.3233/efi-180221.

13. United States Department of Agriculture Natural Resources Conservation Service. Classification for Kingdom Plantae Down to Genus *Ficus* L. Available from: <https://plants.usda.gov/home/classification/70817>. Retrieved October 27, 2024.
14. Rønsted N, Weiblen GD, Cook JM, Salamin N, Machado CA, Savolainen V. 60 million years of co-divergence in the fig-wasp symbiosis. *Proc Biol Sci.* 2005;272(1581):2593-9. doi: 10.1098/rspb.2005.3249.
15. Santos A, Van Ree R. Profilins: mimickers of allergy or relevant allergens? *Int Arch Allergy Immunol.* 2011;155(3):191-204. doi: 10.1159/000321178.
16. Upadhyay RK. Plant latex: a natural source of pharmaceuticals and pesticides: a review. *Int J Green Pharm.* 2011;5(3):169-80.
17. Eisikowitch D, Ghara M. An overview on *Ficus* pollination with some notes on *Ficus carica*. *Italus Hortus.* 2015;22(3):1-7.
18. Marinho CR, Pereira RA, Peng YQ, Teixeira SP. Laticifer distribution in fig inflorescence and its potential role in the fig-fig wasp mutualism. *Acta Oecol (Montrouge).* 2018;90:160-7. doi: 10.1016/j.actao.2017.10.005.
19. Ramya KT, Fiyaz RA, Shaanker RU, Ganeshiah KN. Pollinators for a syconium: how do wasps choose among syconia? *Curr Sci.* 2011;101(4):520-7.
20. Ballaoui M, El Gabardi S, Adnani M, Selmaoui K, Saidi N, Msairi S, et al. Enhanced fig tree production through endomycorrhizal fungi inoculation: a nursery study on tree cuttings derived from the Ifrane region, Morocco. *Not Sci Biol.* 2024;16(3):11972. doi: 10.55779/nsb16311972.
21. Pizon JR, Nuñez OM, Uy MM, Senarath WT. Ethnobotany of medicinal plants used by the Subanen tribe of Lapuyan, Zamboanga del Sur. *Bull Environ Pharmacol Life Sci.* 2016;5(55):53-67.
22. Dapar ML, Alejandro GJ, Meve U, Liede-Schumann S. Quantitative ethnopharmacological documentation and molecular confirmation of medicinal plants used by the Manobo tribe of Agusan del Sur, Philippines. *J Ethnobiol Ethnomed.* 2020;16(1):14. doi: 10.1186/s13002-020-00363-7.
23. Alinsug MV, Estandarte MH, Somodio EM, Sabarita MJ, Deocaris CC. Biodiversity of ethnomedicinal plants from the B'laan tribe in Mount Matutum Protected Landscape, Southern Mindanao, Philippines. *Biodiversitas.* 2022;23(1):554-63. doi: 10.13057/biodiv/d230160.
24. Rosales ER, Casio CR, Amistad VR, Polo CM, Dugaduga KD, Picardal JP. Floristic inventory and ethnobotany of wild edible plants in Cebu Island, Philippines. *Asian J Biodivers.* 2018;9(1):91-115. doi: 10.7828/ajob.v9i1.1236.
25. Agapin JS. Medicinal plants used by traditional healers in Pagadian city, Zamboanga del Sur, Philippines. *Philipp J Sci.* 2020;149(1):83-9. doi: 10.2139/ssrn.3625332.
26. Madjos G, Ramos K. Ethnobotany, systematic review and field mapping on folkloric medicinal plants in the Zamboanga Peninsula, Mindanao, Philippines. *J Complement Med Res.* 2021;12(1):21-61. doi: 10.5455/jcmr.2021.12.01.05.
27. Antonio NC, Tuason RJ. Ethnobotanical and phytochemical study of the medicinal plants used by Kanawan Aytas in Morong, Bataan, Philippines. *Indian J Tradit Knowl.* 2022;21(3):595-604. doi: 10.56042/ijtk.v21i3.31614.
28. Agua AR, Olowa LM. Medicinal Plants Used by the Subanen Tribe in Selected Barangays of Sindangan, Zamboanga del Norte, Philippines. Iligan City, Philippines: Mindanao State University-Iligan Institute of Technology; 2015. p. 1-20.
29. Manua AA. Ethnobotanical Study on the Medicinal Plants Used by the Maranaos in the Municipality of Bubong, Lanao del Sur, Philippines. Iligan City, Philippines: Mindanao State University-Iligan Institute of Technology; 2015. p. 1-32.
30. Olowa L, Demayo CG. Ethnobotanical uses of medicinal plants among the Muslim Maranaos in Iligan City, Mindanao, Philippines. *Adv Environ Biol.* 2015;9(27):204-15.
31. Olowa LF, Torres MA, Aranico EC, Demayo CG. Medicinal plants used by the Higaonon tribe of Rogongon, Iligan City, Mindanao, Philippines. *Adv Environ Biol.* 2012;6(4):1442-9.
32. Haganas JS. Medicinal Plants Used by the Locals of San Jose, Dinagat Islands Province, Philippines. Mindanao State University-Iligan Institute of Technology; 2015. p. 1-28.
33. Wu PL, Rao KV, Su CH, Kuoh CS, Wu TS. Phenanthroindolizidine alkaloids and their cytotoxicity from the leaves of *Ficus septica*. *Heterocycles.* 2002;57(12):2401-8. doi: 10.3987/com-02-9615.
34. Ueda JY, Takagi M, Shin-ya K. Aminocaprophenone- and pyrrolidine-type alkaloids from the leaves of *Ficus septica*. *J Nat Prod.* 2009;72(12):2181-3. doi: 10.1021/np900580f.
35. Yap VA, Qazzaz ME, Raja VJ, Bradshaw TD, Loh HS, Sim KS, et al. Fistulopsines A and B antiproliferative septicine-type alkaloids from *Ficus fistulosa*. *Phytochem Lett.* 2016;15:136-41. doi: 10.1016/j.phytol.2015.12.007.
36. Yap VA, Loong BJ, Ting KN, Loh SH, Yong KT, Low YY, et al. Hispidacine, an unusual 8,4'-oxyneolignan-alkaloid with vasorelaxant activity, and hispiloscine, an antiproliferative phenanthroindolizidine alkaloid, from *Ficus hispida* Linn. *Phytochemistry.* 2015;109:96-102. doi: 10.1016/j.phytochem.2014.10.032.
37. Nugroho AE, Akbar FF, Wiyani A, Sudarsono. Cytotoxic effect and constituent profile of alkaloid fractions from ethanolic extract of *Ficus septica* Burm. F. leaves on T47D breast cancer cells. *Asian Pac J Cancer Prev.* 2015;16(16):7337-42. doi: 10.7314/apjcp.2015.16.16.7337.
38. Al-Khdhairawi AA, Krishnan P, Mai CW, Chung FF, Leong CO, Yong KT, et al. A bis-benzopyrroloisoquinoline alkaloid incorporating a cyclobutane core and a chlorophenanthroindolizidine alkaloid with cytotoxic activity from *Ficus fistulosa* var. *tengerensis*. *J Nat Prod.* 2017;80(10):2734-40. doi: 10.1021/acs.jnatprod.7b00500.
39. Cayetano-Salazar L, de la Cruz-Concepción B, Navarro-Tito N, Álvarez-Fitz P, Leyva-Vázquez MA, Acevedo-Quiroz M, et al. *Ficus crocata* leaf extracts decrease the proliferation and invasiveness of breast cancer cells. *Heliyon.* 2022;8(11):e11405. doi: 10.1016/j.heliyon.2022.e11405.
40. Mustafa K, Yu S, Zhang W, Mohamed H, Naz T, Xiao H, et al. Screening, characterization, and in vitro-ROS dependent cytotoxic potential of extract from *Ficus carica* against hepatocellular (HepG2) carcinoma cells. *S Afr J Bot.* 2021;138:217-26. doi: 10.1016/j.sajb.2020.12.018.
41. Dutta R, Bhattacharya E, Pramanik A, Hughes TA, Mandal Biswas S. Potent nutraceuticals having antioxidant, DNA damage protecting potential and anti-cancer properties from the leaves of four *Ficus* species. *Biocatal Agric Biotechnol.* 2022;44:102461. doi: 10.1016/j.bcab.2022.102461.
42. Zhang Y, Wan Y, Huo B, Li B, Jin Y, Hu X. Extracts and



- components of *Ficus carica* leaves suppress survival, cell cycle, and migration of triple-negative breast cancer MDA-MB-231 cells. *Onco Targets Ther.* 2018;11:4377-86. doi: 10.2147/ott.S171601.
43. Sonibare MA, Sonibare OO, Akhrame OE, Soladoye MO. Chemical composition of essential oils of *Ficus elasticoides* De Wild., *Ficus ovata* Vahl and *Ficus natalensis* subsp. *leprieurii* (Miq.) C.C. Berg from Nigeria. *J Essent Oil Bear Plants.* 2009;12(3):282-6. doi: 10.1080/0972060x.2009.10643721.
  44. Sánchez-Valdeolivar CA, Alvarez-Fitz P, Zacapala-Gómez AE, Acevedo-Quiroz M, Cayetano-Salazar L, Olea-Flores M, et al. Phytochemical profile and antiproliferative effect of *Ficus crocata* extracts on triple-negative breast cancer cells. *BMC Complement Med Ther.* 2020;20(1):191. doi: 10.1186/s12906-020-02993-6.
  45. Soib HH, Yaakob H, Sarmidi MR, Rosdi MN. Fractionation of aqueous extract of *Ficus deltoidea* var. *kunstleri*'s leaves using solid phase extraction method for anticancer activity on du145 cell line. *Malays J Anal Sci.* 2019;23(3):534-47. doi: 10.17576/mjas-2019-2303-18.
  46. De Las Llagas MC, Santiago L, Ramos JD. Cytotoxicity and apoptotic activity of *Ficus pseudopalma* Blanco leaf extracts against human prostate cancer cell lines. *Trop J Pharm Res.* 2014;13(1):93-100. doi: 10.4314/tjpr.v13i1.14.
  47. Van Kiem P, Van Minh C, Nhiem NX, Cuong NX, Tai BH, Quang TH, et al. Inhibitory effect on TNF- $\alpha$ -induced IL-8 secretion in HT-29 cell line by glyceroglycolipids from the leaves of *Ficus microcarpa*. *Arch Pharm Res.* 2012;35(12):2135-42. doi: 10.1007/s12272-012-1210-8.
  48. Parveen M, Malla AM, Alam M, Ahmad F, Silva PS, Silva MR. Two new phenolic compounds from *Ficus rumphii* and their antiproliferative activity. *Nat Prod Res.* 2014;28(9):646-52. doi: 10.1080/14786419.2014.891201.
  49. Prasheena Russell S, Prema Kumari J. Characterization, pharmacology and in-silico study of 2,4 ditertiary butylphenol isolated from the leaves of *Ficus auriculata* Lour. *Orient J Chem.* 2023;39(1):179-88. doi: 10.13005/ojc/390122.
  50. Yalçinkaya T, Kozaci LD, Çarhan A. *Ficus carica* extract causes cell cycle arrest and induces apoptosis in MG-63 and HT-29 cancer cell lines. *Turk Hij Den Biyol Derg.* 2023;80(1):73-88. doi: 10.5505/TurkHijyen.2023.93357.
  51. Alam P, Alhowiriny TA, Siddiqui NA, Alqasoumi SI, Basudan OA, Khan AA, et al. Interspecies estimation of  $\beta$ -sitosterol by a validated high-performance thin-layer chromatography method in genus *Ficus* and cytotoxic activity against HepG2, HEK-293, MCF-7, and MDA-MB-231 cell lines. *Journal of Planar Chromatography Modern TLC.* 2018;31(3):213-9. doi: 10.1556/1006.2018.31.3.6.
  52. Dhawale PG, Ghyare BP. Gas chromatography and mass spectroscopy studies on leaves of *Ficus hispida* L. *Int J Appl Res.* 2016;3(7 Pt D):253-6.
  53. Wangkheirakpam SD, Wadawale A, Leishangthem SS, Gurumayum JS, Laitonjam WS. Cytotoxic triterpenoids from *Ficus pomifera* Wall. *Indian J Chem.* 2015;54(5):676-81.
  54. Shafaei A, Muslim NS, Nassar ZD, Aisha AF, Majid AM, Ismail Z. Antiangiogenic effect of *Ficus deltoidea* Jack standardised leaf extracts. *Trop J Pharm Res.* 2014;13(5):761-8. doi: 10.4314/tjpr.v13i5.16.
  55. Al-Salman HN, Ali ET, Jabir M, Sulaiman GM, Al-Jadaan SA. 2-Benzhydrylsulfinyl-N-hydroxyacetamide-Na extracted from fig as a novel cytotoxic and apoptosis inducer in SKOV-3 and AMJ-13 cell lines via P53 and caspase-8 pathway. *Eur Food Res Technol.* 2020;246(8):1591-608. doi: 10.1007/s00217-020-03515-x.
  56. Zhang J, Zhu WF, Xu J, Kitdamrongtham W, Manosroi A, Manosroi J, et al. Potential cancer chemopreventive and anticancer constituents from the fruits of *Ficus hispida* L.f. (Moraceae). *J Ethnopharmacol.* 2018;214:37-46. doi: 10.1016/j.jep.2017.11.016.
  57. Mbosso EJ, Kamdem LM, Assob JC, Meyer F, Ebelle DC, Lenta BN, et al. In vitro evaluation of antimicrobial and antiproliferative activities for compounds isolated from the *Ficus bubu* Warb. (Moraceae) fruits: chemotaxonomic significance. *Drug Deliv Lett.* 2015;5(2):122-31. doi: 10.2174/2210303105666151008213521.
  58. Maurya A, Priyadarshini E, Rajamani P. Evaluation of antioxidant capacity and antiproliferative activity of fruit extract of dry figs (*Ficus carica* L.) on MDA MB-468 cell line. *Res Sq [Preprint].* December 28, 2021. Available from: <https://www.researchsquare.com/article/rs-1145771/v1>.
  59. Yao J, Wang Z, Wang R, Wang Y, Xu J, He X. Antiproliferative and anti-inflammatory prenylated isoflavones and coumaronochromones from the fruits of *Ficus altissima*. *Bioorg Chem.* 2021;113:104996. doi: 10.1016/j.bioorg.2021.104996.
  60. Bondili PB, Mangamuri UK, Poda S, Mutyala SR, Gali G. Anticancer activity of *Ficus vasta* Forssk. fruit extract in human lung cancer (A549) and human breast cancer (MDA-MB-231) cells. *Int J All Res Educ Sci Methods.* 2023;11(8):458-66.
  61. Al-Shabibi MH, Al-Touby SS, Hossain MA. Isolation, characterization and prediction of biologically active glycoside compounds quercetin-3-rutinoside from the fruits of *Ficus sycomorus*. *Carbohydr Res.* 2022;511:108483. doi: 10.1016/j.carres.2021.108483.
  62. Saini R, Garg V, Dangwal K. Comparative study of three wild edible fruits of Uttarakhand for antioxidant, antiproliferative activities and polyphenolic composition. *Int J Pharma Bio Sci.* 2012;3(4):158-67.
  63. Jing L, Zhang YM, Luo JG, Kong LY. Tirucallane-type triterpenoids from the fruit of *Ficus carica* and their cytotoxic activity. *Chem Pharm Bull (Tokyo).* 2015;63(3):237-43. doi: 10.1248/cpb.c14-00779.
  64. Damu AG, Kuo PC, Shi LS, Li CY, Su CR, Wu TS. Cytotoxic phenanthroindolizidine alkaloids from the roots of *Ficus septica*. *Planta Med.* 2009;75(10):1152-6. doi: 10.1055/s-0029-1185483.
  65. Mbosso EJ, Siwe Noundou X, Nguemfo EL, Meyer F, Djoukoue A, Van Antwerpen P, et al. Identification of compounds with anti-proliferative activity from the wood of *Ficus elastica* Roxb. ex Hornem. aerial roots. *Fitoterapia.* 2016;112:65-73. doi: 10.1016/j.fitote.2016.05.002.
  66. Yen GC, Chen CS, Chang WT, Wu MF, Cheng FT, Shiau DK, et al. Antioxidant activity and anticancer effect of ethanolic and aqueous extracts of the roots of *Ficus beecheyana* and their phenolic components. *J Food Drug Anal.* 2018;26(1):182-92. doi: 10.1016/j.jfda.2017.02.002.
  67. Ye XS, Tian WJ, Liu XZ, Zhou M, Zeng DQ, Lin T, et al. Lignans and phenylpropanoids from the roots of *Ficus hirta* and their cytotoxic activities. *Nat Prod Res.* 2022;36(15):3840-9. doi: 10.1080/14786419.2021.1892099.
  68. Akhtar N, Syed DN, Lall RK, Mirza B, Mukhtar H. Targeting epithelial to mesenchymal transition in prostate

- cancer by a novel compound, plectranthoic acid, isolated from *Ficus microcarpa*. *Mol Carcinog*. 2018;57(5):653-63. doi: 10.1002/mc.22790.
69. Damu AG, Kuo PC, Shi LS, Li CY, Kuoh CS, Wu PL, et al. Phenanthroindolizidine alkaloids from the stems of *Ficus septica*. *J Nat Prod*. 2005;68(7):1071-5. doi: 10.1021/np050095o.
  70. Somwong P, Suttisri R, Buakeaw A. New sesquiterpenes and phenolic compound from *Ficus foveolata*. *Fitoterapia*. 2013;85:1-7. doi: 10.1016/j.fitote.2012.12.026.
  71. Yessoufou K, Elansary HO, Mahmoud EA, Skalicka-Woźniak K. Antifungal, antibacterial and anticancer activities of *Ficus drupacea* L. stem bark extract and biologically active isolated compounds. *Ind Crops Prod*. 2015;74:752-8. doi: 10.1016/j.indcrop.2015.06.011.
  72. Nana F, Sandjo LP, Keumedjio F, Ambassa P, Malik R, Kuete V, et al. Ceramides and cytotoxic constituents from *Ficus glumosa* Del. (Moraceae). *J Braz Chem Soc*. 2012;23(3):482-7. doi: 10.1590/s0103-50532012000300015.
  73. Choudhari AS, Suryavanshi SA, Ingle H, Kaul-Ghanekar R. Evaluating the antioxidant potential of aqueous and alcoholic extracts of *Ficus religiosa* using ORAC assay and assessing their cytotoxic activity in cervical cancer cell lines. *Biotechnol Bioinform Bioeng*. 2011;1(4):443-50.
  74. Popwo Tameye SC, Djamen Mbeunkeu AB, Fouokeng Y, Jouwa Tameye NS, Tabekoueng GB, Wansi JD, et al. Ficusanolide A and ficusanolide B, two new cinnamic acid derivative stereoisomers and other constituents of the stem barks of *Ficus exasperata* Vahl. (Moraceae). *Phytochem Lett*. 2021;43:150-3. doi: 10.1016/j.phytol.2021.03.027.
  75. Tezcan G, Tunca B, Bekar A, Yalcin M, Sahin S, Budak F, et al. *Ficus carica* latex prevents invasion through induction of let-7d expression in GBM cell lines. *Cell Mol Neurobiol*. 2015;35(2):175-87. doi: 10.1007/s10571-014-0109-y.
  76. Tezcan G, Tunca B, Bekar A, Yalcin M, Sahin S, Budak F, et al. *Ficus carica* latex prevents invasion through induction of let-7d expression in GBM cell lines. *Cell Mol Neurobiol*. 2015;35(2):175-87. doi: 10.1007/s10571-014-0109-y.
  77. Hu R, Chantana W, Pitchakarn P, Subhawa S, Chantarasuwan B, Temviriyankul P, et al. *Ficus dubia* latex extract induces cell cycle arrest and apoptosis by regulating the NF- $\kappa$ B pathway in inflammatory human colorectal cancer cell lines. *Cancers (Basel)*. 2022;14(11):2665. doi: 10.3390/cancers14112665.
  78. Jamil EF, Abdul Ghani R. *Ficus auriculata* (fig) extracts induced cell cycle profile changes and apoptosis through caspase-independent pathway in human lung adenocarcinoma cell line, A549. *J Med Plants*. 2017;16(63):57-67.
  79. Chou WM, Chen CN, Hsieh HT, Lo TY, Juan PY, Mai FD. G2/M arrest and apoptosis of human colorectal cancer cells induced by water extract from residues of jelly fig achene. *Technol Health Care*. 2015;24 Suppl 1:S147-53. doi: 10.3233/thc-151063.
  80. Kumar BK, Chary KA, Kiran, Shivani G, Abdul Rahaman SK, Mamatha B. Evaluation of in-vitro cytotoxic potential of *Ficus benghalensis* tender prop roots extract. *Future J Pharm Health Sci*. 2022;2(3):159-69.
  81. Ramakrishna V, Gupt KP, Setty OH, Kondapia AK. Protective effect of *Ficus benghalensis* L. extract against H<sub>2</sub>O<sub>2</sub> induced DNA damage and repair in neuroblastoma cells. *Free Radicals and Antioxidants*. 2014;4(1):3-7. doi: 10.5530/fra.2014.1.2.
  82. Tabrez S, Hoque M, Suhail M, Khan MI, Zughaihi TA, Khan AU. Identification of anticancer bioactive compounds derived from *Ficus* sp. by targeting poly[ADP-ribose]polymerase 1 (PARP-1). *J King Saud Univ Sci*. 2022;34(5):102079. doi: 10.1016/j.jksus.2022.102079.
  83. Yun HJ, Park JH, Kwon HJ, Lee EW, Kim YH, Choi JS, Kim BW. Isolation and identification of a regulatory component that down-regulate MCM protein expression from *Ficus carica*. *J Cancer Prev*. 2008;13(4):302-10.
  84. AlGhalban FM, Khan AA, Khattak MN. Comparative anticancer activities of *Ficus carica* and *Ficus salicifolia* latex in MDA-MB-231 cells. *Saudi J Biol Sci*. 2021;28(6):3225-34. doi: 10.1016/j.sjbs.2021.02.061.
  85. Soltana H, Pinon A, Limami Y, Zaid Y, Khalki L, Zaid N, et al. Antitumoral activity of *Ficus carica* L. on colorectal cancer cell lines. *Cell Mol Biol (Noisy-le-grand)*. 2019;65(6):6-11.
  86. Khodarahmi GA, Ghasemi N, Hassanzadeh F, Safaie M. Cytotoxic effects of different extracts and latex of *Ficus carica* L. on HeLa cell Line. *Iran J Pharm Res*. 2011;10(2):273-7.
  87. Hamid IS, Aksono EB, Sukmanadi M, Purnama MT. Antiangiogenesis activity test of tin leaf (*Ficus carica* L.) on the number of blood vessels and VEGF expression of chorioallantoic membrane of embryonated chicken eggs. *Eur J Oncol Pharm*. 2018;1(4):e00007. doi: 10.1097/op9.0000000000000007.
  88. Dundar T, Kocyigit A, Guler EM. Investigation of *Ficus carica* plantae' cytotoxic, genotoxic, apoptotic, antineoplastic, anti-inflammatory, and autophagic effects on breast cancer cells. *Bezmialem Sci*. 2021;9(S2):S24.
  89. Lee SA, Park BR, Kim CS. Induction of apoptosis by methanol extracts of *Ficus carica* L. in FaDu human hypopharynx squamous carcinoma cells. *Int J Oral Biol*. 2020;45(3):99-106. doi: 10.11620/ijob.2020.45.3.99.
  90. Abolmaesoomi M, Abdul Aziz A, Mat Junit S, Mohd Ali J. *Ficus deltoidea*: effects of solvent polarity on antioxidant and anti-proliferative activities in breast and colon cancer cells. *Eur J Integr Med*. 2019;28:57-67. doi: 10.1016/j.eujim.2019.05.002.
  91. Akhir NA, Chua LS, Majid FA, Sarmidi MR. Cytotoxicity of aqueous and ethanolic extracts of *Ficus deltoidea* on human ovarian carcinoma cell line. *Br J Med Med Res*. 2011;1(4):397-409. doi: 10.9734/bjmmr/2011/507.
  92. Al-Koshab M, Alabsi AM, Mohd Bakri M, Ali-Saeed R, Selvi Naicker M. Antitumor activity of *Ficus deltoidea* extract on oral cancer: an in vivo study. *J Oncol*. 2020;2020:5490468. doi: 10.1155/2020/5490468.
  93. Mahmoud K, Khalil WK, Elmoniem OA, Mahrous KF. Evaluation of *Ficus glomerata* extract as potential anticancer agents and prevents the genetic toxicity induced by benzo(a)pyrene in male mice. *Int J PharmTech Res*. 2015;8(4):678-90.
  94. Vakili S, Pasha MJ. Phytochemical investigation and cytotoxic screening of three medicinal plants which are traditionally used in treating cancer in Ayurvedic medicine. *Saudi Pharm J*. 2016;10:1185-204.
  95. Zeng YW, Liu XZ, Lv ZC, Peng YH. Effects of *Ficus hirta* Vahl. (Wuzhimaotao) extracts on growth inhibition of HeLa cells. *Exp Toxicol Pathol*. 2012;64(7-8):743-9. doi: 10.1016/j.etp.2011.01.009.
  96. Sathiyamoorthy J, Sudhakar N. In vitro cytotoxicity and apoptotic assay in HT-29 cell line using *Ficus hispida* Linn: leaves extract. *Pharmacogn Mag*. 2018;13(Suppl 4):S756-61. doi: 10.4103/pm.pm\_319\_17.



97. Kanjkar AP, Aruna LH, Londonkar RL. Novel efficacy of in vitro anti-haemolytic and anti-cancer activities of *Ficus krishnae*. *Pharm Lett.* 2017;9(12):16-22.
98. Akhtar N, Syed DN, Khan MI, Adhami VM, Mirza B, Mukhtar H. The pentacyclic triterpenoid, plectranthoic acid, a novel activator of AMPK induces apoptotic death in prostate cancer cells. *Oncotarget.* 2016;7(4):3819-31. doi: 10.18632/oncotarget.6625.
99. Lv H, Hu C, Xie Z, Wang P, Chen X, Wen C. Purification, characterization and anti-tumor activity of a pectic-type polysaccharide isolated from *Ficus pandurata* H. *Int J Biol Macromol.* 2020;153:201-6. doi: 10.1016/j.ijbiomac.2020.02.244.
100. Santiago LA, Mayor AB. Antiproliferative and apoptotic effects of *Ficus pseudopalma* Blanco (Moraceae) against hepatocarcinoma (HepG2). *Asian J Pharm Clin Res.* 2015;8(2):257-61.
101. Le TT, Nguyen MQ, Nguyen VP, Hoang VH, Nguyen PH. Methanol extract of *Ficus pumila* L. inhibits proliferation, induces apoptosis and arrests the cell cycle in HepG2 live cancer cells. *Asian J Plant Sci.* 2023;22(3):423-33. doi: 10.3923/ajps.2023.423.433.
102. Syahputra HD, Masfria M, Hasibuan PA, Iksen I. In silico docking studies of phytoesterol compounds selected from *Ficus religiosa* as potential chemopreventive agent. *Rasayan J Chem.* 2022;15(2):1080-4.
103. Haneef J, Parvathy M, Thankayyan RS, Sithul H, Sreeharshan S. Bax translocation mediated mitochondrial apoptosis and caspase dependent photosensitizing effect of *Ficus religiosa* on cancer cells. *PLoS One.* 2012;7(7):e40055. doi: 10.1371/journal.pone.0040055.
104. Maity A, Bisoi PC, Behera PC, Sahoo GR. Cytotoxicity study on *Ficus religiosa* and *Ziziphus mauritiana* leaf extracts against MCF-7 cancer cell line. *Indian Journal of Field Veterinarians.* 2011;7(1):47-8.
105. Maity A, Bisoi PC, Das AB, Maiti P, Senapati MR. Cytotoxicity study of *Ficus religiosa* leaf extract against Jurkat cell line. *Explor Anim Med Res.* 2017;7(1):18-21.
106. Nugroho AE, Ikawati M, Hermawan A, Putri DD, Meiyanto E. Cytotoxic effect of ethanolic extract fractions of Indonesia plant *Ficus septica* Burm. F. on human breast cancer T47D cell lines. *Int J Phytomed.* 2011;3(2):216-26. doi: 10.5138/ijpm.v3i2.331.
107. Anindyajati A, Darma AP, Nurjizah I, Septhea DB, Nugroho AE. *Ficus septica* Burm. F. leaves ethanolic extract triggered apoptosis on 7,12-dimethylbenz[a]anthracene-induced rat mammary carcinogenesis qualitatively. *Indones J Cancer Chemoprevention.* 2012;3(1):334-8. doi: 10.14499/indonesianjcanchemprev3iss1pp334-338.
108. Canoy RJ, Lomanta JM, Ballesteros PM, Chun EA, Dator RP, Jacinto SD. Cancer chemotherapeutic potential of endemic and indigenous plants of Kanawan, Morong, Bataan province, Philippines. *Asia Life Sci.* 2010;20(2):331-9.
109. Krupanidhi S, Madhan Sai N, Leung H, Kineman JJ. The leaf as a sustainable and renewable system. *Syst Res Behav Sci.* 2017;34(5):564-76. doi: 10.1002/sres.2487.
110. Morilla LJ, Sumaya NH, Rivero HI, Madamba MR. Medicinal plants of the Subanans in Dumingag, Zamboanga del Sur, Philippines. *Int Conf Food Biol Med Sci.* 2014;10:38-43. doi: 10.15242/iicbe.c0114577.
111. Cordero C, Alejandro GJ. Medicinal plants used by the indigenous Ati tribe in Tobias Fornier, Antique, Philippines. *Biodiversitas.* 2021;22(2):521-36. doi: 10.13057/biodiv/d220203.
112. Rubio MM, Naïve MA. Ethnomedicinal plants used by traditional healers in North Cotabato, Mindanao, Philippines. *J Biodivers Environ Sci.* 2018;13(6):74-82.
113. Jha AK, Sit N. Extraction of bioactive compounds from plant materials using combination of various novel methods: a review. *Trends Food Sci Technol.* 2022;119:579-91. doi: 10.1016/j.tifs.2021.11.019.
114. Bucjan ER. Descriptive analysis of folk narratives of the Kamayo in Surigao del Sur, Philippines. *SDSSU Multidiscip Res J.* 2019;7:5-9.
115. Berdon JS, Ragosta EL, Inocian RB, Manalag CA, Lozano EB. Unveiling Cebuano traditional healing practices. *Asia Pac J Multidiscip Res.* 2016;4(1):51-9.
116. Simbulan NP. The health perceptions and practices of Lumads in Southern Philippines. *Acta Med Philipp.* 2011;45(2):37-49.
117. Sosa V, Moliné T, Somoza R, Paciucci R, Kondoh H, Lleonart ME. Oxidative stress and cancer: an overview. *Ageing Res Rev.* 2013;12(1):376-90. doi: 10.1016/j.arr.2012.10.004.
118. Hayes JD, Dinkova-Kostova AT, Tew KD. Oxidative stress in cancer. *Cancer Cell.* 2020;38(2):167-97. doi: 10.1016/j.ccell.2020.06.001.
119. Villard C, Larbat R, Munakata R, Hehn A. Defence mechanisms of *Ficus*: pyramiding strategies to cope with pests and pathogens. *Planta.* 2019;249(3):617-33. doi: 10.1007/s00425-019-03098-2.
120. Roaa MH. A review article: the importance of the major groups of plants secondary metabolism phenols, alkaloids, and terpenes. *Int J Res Appl Sci Biotechnol.* 2020;7(5):354-8.
121. Oliveira AP, Silva LR, Guedes de Pinho P, Gil-Izquierdo A, Valentão P, Silva BM, et al. Volatile profiling of *Ficus carica* varieties by HS-SPME and GC-IT-MS. *Food Chem.* 2010;123(2):548-57. doi: 10.1016/j.foodchem.2010.04.064.
122. Luo SH, Hua J, Liu Y, Li SH. The chemical ecology of plant natural products. *Prog Chem Org Nat Prod.* 2024;124:57-183. doi: 10.1007/978-3-031-59567-7\_2.
123. Shen N, Wang T, Gan Q, Liu S, Wang L, Jin B. Plant flavonoids: classification, distribution, biosynthesis, and antioxidant activity. *Food Chem.* 2022;383:132531. doi: 10.1016/j.foodchem.2022.132531.
124. Di Ferdinando M, Brunetti C, Fini A, Tattini M. Flavonoids as antioxidants in plants under abiotic stresses. In: Ahmad P, Prasad MN, eds. *Abiotic Stress Responses in Plants: Metabolism, Productivity and Sustainability.* New York, NY: Springer; 2012. p. 159-79. doi: 10.1007/978-1-4614-0634-1\_9.
125. Shah A, Smith DL. Flavonoids in agriculture: chemistry and roles in, biotic and abiotic stress responses, and microbial associations. *Agronomy.* 2020;10(8):1209. doi: 10.3390/agronomy10081209.
126. Ferreyra ML, Serra P, Casati P. Recent advances on the roles of flavonoids as plant protective molecules after UV and high light exposure. *Physiol Plant.* 2021;173(3):736-49. doi: 10.1111/ppl.13543.
127. Vinson JA, Zubik L, Bose P, Samman N, Proch J. Dried fruits: excellent in vitro and in vivo antioxidants. *J Am Coll Nutr.* 2005;24(1):44-50. doi: 10.1080/07315724.2005.10719442.
128. Zhao D, Tao J. Recent advances on the development and regulation of flower color in ornamental plants. *Front Plant Sci.* 2015;6:261. doi: 10.3389/fpls.2015.00261.

129. Huang X, Gao S, Fan L, Yu S, Liang X. Cytotoxic alkaloids from the roots of *Tylophora atrofoliculata*. *Planta Med.* 2004;70(5):441-5. doi: 10.1055/s-2004-818973.
130. Tajner-Czopek A, Gertchen M, Rytel E, Kita A, Kucharska AZ, Sokół-Łętowska A. Study of antioxidant activity of some medicinal plants having high content of caffeic acid derivatives. *Antioxidants (Basel).* 2020;9(5):412. doi: 10.3390/antiox9050412.
131. Taghizadeh M, Soltanian S, Nasibi N. Phytochemical analysis of volatile and non-volatile fractions, antioxidant, and anti-cancer activities of *Dracocephalum polychaetum* and *Dracocephalum kotschyi*. *J Cell Mol Res.* 2022;14(1):11-9. doi: 10.22067/jcmr.2022.75035.1031.
132. Salomé Abarca LF, Klinkhamer PGL, Choi YH. Plant latex, from ecological interests to bioactive chemical resources. *Planta Med.* 2019;85(11-12):856-68. doi: 10.1055/a-0923-8215.
133. Sun W, Shahrajabian MH. Therapeutic potential of phenolic compounds in medicinal plants-natural health products for human health. *Molecules.* 2023;28(4):1845. doi: 10.3390/molecules28041845.
134. Keshari AK, Kumar G, Kushwaha PS, Bhardwaj M, Kumar P, Rawat A, et al. Isolated flavonoids from *Ficus racemosa* stem bark possess antidiabetic, hypolipidemic and protective effects in albino Wistar rats. *J Ethnopharmacol.* 2016;181:252-62

**Copyright** © 2025 The Author(s). This is an open-access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.