Aqueous extract of *Combretum molle* boughs ameliorates hyperglycaemia and hyperlipidemia in sucrose-induced insulin resistant rats

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**Implication for health policy/practice/research/medical education:**
The aqueous extract of *Combretum molle* boughs may be able to delay onset of insulin resistance, and reduce the risks and complications of type 2 diabetes. Therefore, its use in diabetic patients is recommended.


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**Abstract**

**Introduction:** *Combretum molle* R.B/G.Don (Combretaceae) is distributed especially in tropical Africa and used in treatment various diseases including diabetes. The aim of the present study was to evaluate the effects of aqueous extract from *C. molle* boughs (CMAE) on hyperglycemia and dyslipidemia in insulin resistant rats.

**Methods:** Animals were divided into 5 groups and treated for 30 days. Control group received distilled water, sucrose group received 30% sucrose, standard group received 30% sucrose plus metformin (40 mg/kg), and others groups received 30% sucrose plus CMAE (250 and 500 mg/kg). Body weight, food and water intake were evaluated each 10 days for 30 days. Glucose tolerance test was performed on the 30th day of the experiment. Later on, animals were sacrificed and blood was collected for the determination of the concentration of glucose, lipids and insulin.

**Results:** The body weight and food intake of the rats receiving 500 mg/kg of extract decreased significantly on the 30th day of the experiment. CMAE caused a significant reduction of insulin, glucose, cholesterol, triglycerides and low-density lipoprotein cholesterol levels compared to the sucrose lot. However, the extract (250 and 500 mg/kg) showed a significant increase in high-density lipoprotein cholesterol. CMAE induced a significant decrease in postprandial glycaemia.

**Conclusion:** CMAE improved postprandial hyperglycaemia and hyperlipidemia in insulin resistant rats. Consequently, CMAE may be able to delay onset of insulin resistance, and reduce the risks and complications of type 2 diabetes.

**Introduction**

Insulin resistance is associated with a decrease in insulin sensitivity in the target tissues (liver, muscles and adipose tissue) (1). Insulin resistance is at the heart of the metabolic syndrome and appears to be a major element of many pathologies such as diabetes, high blood pressure, heart and vessel diseases (2). Thus, diabetes mellitus is a public health problem because all these diseases are increasing worldwide, both in industrialized and developing countries. Indeed, globally, approximately 422 million diabetics were identified in 2014 and by 2030, about 430 million people will have this disease (3). Diabetes deaths were 1.5 million in 2012 and will reach 3.7 million in 2040 (4).

High-sucrose intake was shown to contribute syndromes such as hyperlipidemia, glucose intolerance, hypertension,
and cardiovascular complications (5). A diet too rich in sucrose can reduce the sensitivity of target tissues to insulin and therefore can induce type 2 diabetes mellitus in animals (6). The treatment of diabetes relies on the administration of insulin and the taking of oral antidiabetic agents such as metformin, sulphonylureas, glinides, alpha-glucosidase inhibitors, etc. These medications have many adverse side effects (hypoglycemia, coma, ketoadiposis, gastrointestinal disorders, headache, nausea, etc.) and significant risks of cardiovascular diseases (7).

The traditional pharmacopoeia offers an alternative solution to most diseases (8). Nowadays, different parts of plants are traditionally used to treat diabetic patients. *Combretum molle* (Combretaceae) is a graceful deciduous shrub 3-13 m in height (9), generally widespread in tropical Africa (10) and widely used in African traditional medicine as antifungal, antimicrobial, antioxidant, anti-inflammatory and antidiabetic (11,12). A literature review revealed no studies of the hypoglycemic activity of the aqueous extract of CMAE boughs in high sucrose-fed experimental animals. The aim of the present study was to explore the activity of CMAE on hyperglycemia and hyperlipidemia in sucrose-induced insulin resistance rats.

### Materials and Methods

**Chemicals**

Kits for biochemical assays were purchased from Sigma-Aldrich (St. Louis, USA). Sucrose and D-glucose were purchased from Edu-Lab Biology Kit (Bexwell, Norfolk, UK). All chemicals and drugs were obtained commercially in analytical grade.

**Plant material and extraction**

The fresh boughs of *C. molle* were collected in December 2018 from Moutourwa, Cameroon. Botanical identification was carried out at the National Herbarium, Yaoundé, Cameroon (HNC). Voucher specimen was maintained in the HNC and registered under N° 433724NHC. The boughs of *C. molle* were washed, chopped into small pieces, dried under shade and finely powdered. Two hundred grams (200 g) of fine powder of *C. molle* were added to 500 mL of distilled water and the whole content was boiled for 15 minutes. After cooling, the crude extract obtained was filtered using Whatman paper and the filtrate was evaporated in an oven for 72 hours at a temperature of 45°C to give a yield of 9.64% of dry extract.

**Experimental animals**

Male Wistar strain rats (220 and 250 g) were provided by the animal house of the Department of Animal Biology of the Faculty of Science at the University of Dschang in the polystyrene cages (5 rats per cage). Before being used for the various tests, they were acclimated for 7 days to laboratory conditions (24 ± 2° C, 50-55% humidity and 12 h light/dark cycles). They received daily drinking water and normal laboratory standard pelleted diet ad libitum.

**Experimental design**

Thirty rats were distributed into 5 different treatment groups (n = 6) for 30 days as follows: Control normal group received distilled water, sucrose group received 30% sucrose solution (Suc), standard group received 30% sucrose plus metformin (Suc + Met) at doses of 40 mg/kg, and others groups received 30% sucrose plus CMAE (Suc + Ext) at doses of 250 and 500 mg/kg. Sucrose solution was administered as drinking water and treatments (either vehicle, standard drug or extract) were administered orally using intra gastric tube. Food, water intake and body weight were evaluated on 0, 10th, 20th and 30th days of the experiment.

**Oral glucose tolerance test (OGTT)**

The glucose tolerance test was performed on the last day of the experiment. Animals were fasted for 12 h and D-glucose solution (2 g/kg) was orally given to the rats. Glycaemia was determined every 30 minutes for 2 hours after administration of D-glucose from tail vein.

**Biochemical estimation**

After the OGTT, animals were fasted overnight and the blood was obtained by decapitation of rats anesthetized with diazepam (10 mg/kg bw, i.p.) and ketamine (50 mg/kg bw, i.p.). Dry tubes containing collected blood were centrifuged at 3000 rpm for 15 min. The obtained serum was stored at -20°C for the estimation of biochemical parameters. Fasting blood glucose was evaluated at the end of the experiment. Serum insulin was measured using ELISA kit. HOMA-IR was calculated as follows: [Serum insulin (µg/L) × Blood glucose (mg/dL)]/22.5. Total cholesterol (TC), triglycerides (TG), high-density lipoprotein cholesterol (HDL-C) and low-density lipoprotein cholesterol (LDL-C) were performed as per the respective kit inserts.

**Statistical analysis**

All data were presented as mean ± SEM. Statistical analysis was carried out by one-way ANOVA followed by Tukey’s post hoc using GraphPad Prism version 5.0. Values of P < 0.05 were considered significant.

### Results

**Effect of CMAE on body weight, food and water consumption**

Table 1 shows the effect of repeated administration of CMAE on body weight, food and water consumption in insulin resistance rats. There was no important variation between the body weight, food and water intake of the control, standard and sucrose groups rats for 30 days of experiment. Body weight decreased significantly in rats.
receiving 500 mg/kg of extract at days 20 (P<0.01) and 30 (P<0.05) of the experiment, compared with untreated insulin resistant rats. Similarly, at the end of the treatment period (day 30), animals treated with 250 and 500 mg/kg of extract showed a significant (P<0.01) reduction in dietary intake. However, water consumption was not different significantly in all experimental animals for 30 days.

**Effect of CMAE on glycaemia, insulin level and HOMA-IR index**

As shown in Table 2, a significant (P<0.001) increase in blood glucose, insulin level and HOMA-IR index was noted in the sucrose group compared to the normal control group. Administration of metformin (P<0.01) and doses of 250 (P<0.01) and 500 mg/kg (P<0.001) of CMAE resulted in a significant decrease in blood glucose compared to insulin resistant group. In addition, a significant (P<0.001) decrease in serum insulin level and HOMA-IR was observed in rats receiving standard drug and different doses of CMAE. In addition, a significant (P<0.01) decrease in serum insulin and HOMA-IR was recorded in rats receiving the standard drug and the different doses of CMAE.

**Oral glucose tolerance test in insulin resistant animals**

The results shown in Table 3 indicate that, compared to the normal control group, administration of sucrose to normal rats resulted in a significant (P<0.001) increase in blood glucose for 2 hours after D-glucose administration. Oral administration of metformin revealed a significant decline in glucose level at 60 (P<0.01), 90 (P<0.001) and 120 min (P<0.001) compared to disease control group. In addition, different doses of CMAE produced an important (P<0.001) decrease in glycaemia at 60, 90 and 120 min after the glucose load.

**Effect of CMAE on lipid parameters**

Sucrose significantly (P<0.001) increased TC, TG and LDL-c levels, and reduced HDL-c level compared to the normal control group (Table 4). Rats treated with 250 and 500 mg/kg of CMAE and metformin showed a significant (P<0.001) decrease in TC, TG and LDL-c levels compared with untreated insulin resistance control. However, a significant increase in HDL-c was recorded in rats receiving the standard drug (P<0.05) and CMAE.

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**Table 1. Effect of Combretum molle on body weight, water and food consumption in insulin resistant rats**

<table>
<thead>
<tr>
<th>Groups</th>
<th>Time (days)</th>
<th>Normal control</th>
<th>Disease control</th>
<th>Suc + Met 40 mg/kg</th>
<th>Suc + Ext 250 mg/kg</th>
<th>Suc + Ext 500 mg/kg</th>
</tr>
</thead>
<tbody>
<tr>
<td>Body weight (g)</td>
<td>0</td>
<td>231.00 ± 4.43</td>
<td>232.18 ± 4.39</td>
<td>231.33 ± 3.26</td>
<td>233.34 ± 4.06</td>
<td>235.33 ± 3.95</td>
</tr>
<tr>
<td></td>
<td>10</td>
<td>238.67 ± 3.99</td>
<td>239.66 ± 4.81</td>
<td>241.83 ± 3.39</td>
<td>242.82 ± 3.67</td>
<td>244.00 ± 4.15</td>
</tr>
<tr>
<td></td>
<td>20</td>
<td>247.67 ± 4.10</td>
<td>251.16 ± 6.03</td>
<td>250.34 ± 2.78</td>
<td>249.17 ± 3.20</td>
<td>248.17 ± 3.97b</td>
</tr>
<tr>
<td></td>
<td>30</td>
<td>250.00 ± 4.07</td>
<td>252.67 ± 5.91</td>
<td>252.50 ± 3.11</td>
<td>251.33 ± 3.59</td>
<td>251.50 ± 4.04a</td>
</tr>
<tr>
<td>Food intake (g)</td>
<td>0</td>
<td>18.92 ± 0.37</td>
<td>20.25 ± 0.66</td>
<td>20.16 ± 0.47</td>
<td>19.48 ± 0.52</td>
<td>18.40 ± 1.43</td>
</tr>
<tr>
<td></td>
<td>10</td>
<td>18.78 ± 0.97</td>
<td>18.02 ± 1.00</td>
<td>16.90 ± 0.97</td>
<td>16.22 ± 1.11</td>
<td>16.52 ± 1.06</td>
</tr>
<tr>
<td></td>
<td>20</td>
<td>18.68 ± 0.78</td>
<td>17.20 ± 1.11</td>
<td>18.58 ± 0.96</td>
<td>16.45 ± 1.05</td>
<td>16.18 ± 0.93</td>
</tr>
<tr>
<td></td>
<td>30</td>
<td>16.58 ± 0.97</td>
<td>19.55 ± 1.00</td>
<td>17.07 ± 0.79</td>
<td>15.07 ± 0.79b</td>
<td>15.72 ± 1.01b</td>
</tr>
<tr>
<td>Water intake (mL)</td>
<td>0</td>
<td>41.31 ± 2.17</td>
<td>42.55 ± 3.78</td>
<td>40.63 ± 3.54</td>
<td>40.42 ± 2.66</td>
<td>37.02 ± 2.50</td>
</tr>
<tr>
<td></td>
<td>10</td>
<td>39.57 ± 4.11</td>
<td>36.48 ± 2.93</td>
<td>37.55 ± 2.46</td>
<td>37.88 ± 2.60</td>
<td>32.20 ± 1.11</td>
</tr>
<tr>
<td></td>
<td>20</td>
<td>40.78 ± 2.48</td>
<td>38.77 ± 1.65</td>
<td>37.20 ± 3.00</td>
<td>35.60 ± 2.68</td>
<td>35.65 ± 3.22</td>
</tr>
<tr>
<td></td>
<td>30</td>
<td>40.32 ± 3.22</td>
<td>38.68 ± 2.04</td>
<td>38.60 ± 3.31</td>
<td>32.88 ± 2.87</td>
<td>35.87 ± 2.72</td>
</tr>
</tbody>
</table>

Data are mean ± SEM (n = 6 in each group). *P<0.05; **P<0.01 significantly different compared to disease control group. Sucrose + Metformin (Suc + Met); Sucrose + Extract (Suc + Ext).

**Table 2. Effect of Combretum molle on blood glucose level, serum insulin and HOMA-IR index in insulin resistant rats**

<table>
<thead>
<tr>
<th>Groups</th>
<th>Time</th>
<th>Normal control</th>
<th>Disease control</th>
<th>Suc + Met 40 mg/kg</th>
<th>Suc + Ext 250 mg/kg</th>
<th>Suc + Ext 500 mg/kg</th>
</tr>
</thead>
<tbody>
<tr>
<td>Glucose (mg/dL)</td>
<td>0</td>
<td>94.33 ± 2.87</td>
<td>118.67 ± 2.95***</td>
<td>97.83 ± 4.76*</td>
<td>99.17 ± 1.72c</td>
<td>91.67 ± 2.81c</td>
</tr>
<tr>
<td></td>
<td>10</td>
<td>250.34 ± 3.59</td>
<td>37.88 ± 2.60</td>
<td>35.60 ± 2.68</td>
<td>35.65 ± 3.22</td>
<td>35.87 ± 2.72</td>
</tr>
<tr>
<td>Insulin (µg/L)</td>
<td>0</td>
<td>0.24 ± 0.00</td>
<td>1.01 ± 0.04***</td>
<td>0.68 ± 0.04***</td>
<td>0.45 ± 0.06**c</td>
<td>0.22 ± 0.03fc</td>
</tr>
<tr>
<td></td>
<td>10</td>
<td>1.95 ± 0.24**c</td>
<td>0.89 ± 0.11c</td>
<td>0.22 ± 0.03fc</td>
<td>0.89 ± 0.11c</td>
<td>0.22 ± 0.03fc</td>
</tr>
<tr>
<td>HOMA-IR</td>
<td>0</td>
<td>0.99 ± 0.03</td>
<td>5.31 ± 0.22***</td>
<td>2.93 ± 0.04***</td>
<td>1.95 ± 0.24**c</td>
<td>0.89 ± 0.11c</td>
</tr>
<tr>
<td></td>
<td>10</td>
<td>38.80 ± 2.72</td>
<td>37.02 ± 2.50</td>
<td>24.91 ± 3.20</td>
<td>24.87 ± 3.97b</td>
<td>24.87 ± 3.97b</td>
</tr>
</tbody>
</table>

Data are the mean ± SEM (n = 6 in each group). *P<0.05; **P<0.01 significantly different compared to normal control group. P<0.01; P<0.001 significantly different compared to disease control group. Sucrose + Metformin (Suc + Met); Sucrose + Extract (Suc + Ext); Homeostatic Model Assessment of Insulin Resistance (HOMA-IR).
Combretum molle improves hyperglycemia and hyperlipidemia in insulin resistance rats

Table 3. Effect of Combretum molle on oral glucose tolerance test in insulin resistant rats

<table>
<thead>
<tr>
<th>Time (min)</th>
<th>Normal control</th>
<th>Disease control</th>
<th>Suc + Met 40 mg/kg</th>
<th>Suc + Ext 250 mg/kg</th>
<th>Suc + Ext 500 mg/kg</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>94.33 ± 2.87</td>
<td>127.83 ± 9.4</td>
<td>122.00 ± 3.30</td>
<td>117.50 ± 3.08</td>
<td>110.17 ± 3.05</td>
</tr>
<tr>
<td>30</td>
<td>118.66 ± 2.95</td>
<td>156.33 ± 3.04</td>
<td>151.50 ± 2.03</td>
<td>151.00 ± 1.88</td>
<td>145.67 ± 1.84</td>
</tr>
<tr>
<td>60</td>
<td>97.83 ± 4.76</td>
<td>131.83 ± 3.52</td>
<td>121.67 ± 2.75a</td>
<td>109.50 ± 2.35b</td>
<td>103.33 ± 4.57c</td>
</tr>
<tr>
<td>90</td>
<td>99.17 ± 1.72</td>
<td>133.83 ± 2.69</td>
<td>124.83 ± 2.75a</td>
<td>119.17 ± 2.02c</td>
<td>111.17 ± 3.52c</td>
</tr>
<tr>
<td>120</td>
<td>91.67 ± 2.81</td>
<td>121.67 ± 2.56</td>
<td>127.83 ± 3.94c</td>
<td>108.17 ± 3.03c</td>
<td>100.83 ± 2.79c</td>
</tr>
</tbody>
</table>

Data are the mean ± SEM (n = 6 in each group). ***p < 0.001 significantly different compared to normal control group. *p < 0.01; **p < 0.001; ***p < 0.0001 significantly different compared to disease control group. Sucrose + Metformin (Suc + Met), Sucrose + Extract (Suc + Ext).

Table 4. Effect of Combretum molle on lipid parameters in insulin resistant animals

<table>
<thead>
<tr>
<th>Groups</th>
<th>Normal control</th>
<th>Disease control</th>
<th>Suc + Met 40 mg/kg</th>
<th>Suc + Ext 250 mg/kg</th>
<th>Suc + Ext 500 mg/kg</th>
</tr>
</thead>
<tbody>
<tr>
<td>TC (mg/dL)</td>
<td>104.53 ± 5.57</td>
<td>210.55 ± 5.06***</td>
<td>157.86 ± 5.08b</td>
<td>167.77 ± 9.93c</td>
<td>152.32 ± 7.03c</td>
</tr>
<tr>
<td>TG (mg/dL)</td>
<td>90.01 ± 3.97</td>
<td>134.79 ± 3.94***</td>
<td>109.38 ± 4.44a</td>
<td>92.54 ± 6.32a</td>
<td>97.51 ± 3.99c</td>
</tr>
<tr>
<td>HDL-C (mg/dL)</td>
<td>28.32 ± 2.16</td>
<td>11.51 ± 1.45***</td>
<td>21.52 ± 3.30b</td>
<td>24.43 ± 2.50b</td>
<td>23.65 ± 2.40a</td>
</tr>
<tr>
<td>LDL-C (mg/dL)</td>
<td>58.21 ± 5.21</td>
<td>172.08 ± 4.65***</td>
<td>114.46 ± 3.96c</td>
<td>124.83 ± 10.79c</td>
<td>109.16 ± 6.10a</td>
</tr>
</tbody>
</table>

Data are mean ± SEM (n = 6 in each group). ***p < 0.001 significantly different compared to normal control group. *p < 0.01; **p < 0.001; ***p < 0.0001 significantly different compared to disease control group. Sucrose + Metformin (Suc + Met), Sucrose + Extract (Suc + Ext), Triglycerides (TG), Total cholesterol (TC), High-density lipoprotein cholesterol (HDL-C), Low-density lipoprotein cholesterol (LDL-C).

Discussion

Many studies have shown that a diet too rich in sucrose causes insulin resistance in rodents (5,13,14), and therefore may affect the sugar and insulin metabolism (15). The mechanisms by which high fat and/or high sucrose diets cause insulin resistance are not fully understood. It has been shown that high sucrose diets may affect the metabolism of insulin and glucose. Preliminary phytochemical studies of aqueous extracts of CMAE showed presence of saponins, phenols, tannins, glycosides, flavonoids, and terpenoids (16). In this work, administration of sucrose in normal rats resulted in a significant increase in blood glucose, glucose intolerance, serum insulin, HOMA-IR, and lipid parameters.

The results showed that the body weight of the animals receiving the dose of 500 mg/kg of CMAE decreased significantly at the 20th and 30th days of treatment. This decrease in body weight could be due to the presence in the extract of chemical compounds such as tannins and saponins. It is documented that tannins have antinutritional properties and therefore, they can lead to loss of body weight, either by complexing proteins in the intestinal lumen (17) or by reducing food consumption (18). Significant decrease in food consumption observed at the last day of treatment in animals receiving different doses of extract partly confirms the observed weight loss.

In this study, CMAE significantly reduced blood glucose, serum insulin, and HOMA-IR compared to untreated sick rats. According to the results of our study, CMAE (250 and 500 mg/kg) may enhance glucose intake in tissues, improve insulin resistance, increase insulin sensitivity and glucose uptake in rats in which insulin resistance was induced by a sucrose solution (19).

On the other hand, different doses of CMAE significantly decreased glycaemia from the 60th min after glucose load. These results suggest that CMAE can improve glucose intolerance and to effectively regulate postprandial glucose.

The oral administration of high sucrose in normal rats led to the dyslipidemia, characterized by high plasma concentrations of TC, TG and LDL-c with low plasma levels of HDL-c (20,21). Results of this study indicate that, CMAE administered during 30 days reduced TC, TG and LDL-c, and increased serum concentration of HDL-c. CMAE could therefore potentially reduce cardiovascular complications under insulin resistance conditions. Moreover, since hyperlipidemia interferes with the absorption of sugar in the muscles, CMAE could stimulate the absorption of glucose by this muscle tissue (22).

Glycosides, flavonoids and phenols in plants would reduce hyperglycemia and hyperlipidemia in insulin-resistant rats (23). The effects of insulin sensitization, anti-hyperglycemia, and anti-hyperlipidemia noted in this study may be due to the presence of these chemicals in the CMAE.

Conclusion

From this study, we can conclude that CMAE has insulin sensitizing and anti-hyperlipidemia potentials in high sucrose-induced insulin resistant rats. CMAE may prevent target tissue resistance to insulin action and thus reduce the risk of developing type 2 diabetes and its cardiovascular complications.
Acknowledgement
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Authors’ contributions
GT helped harvest and identification of plant material. AD proposed plant material and provided the reagents for the realization of phytochemical screening. DM and AD performed the crude extract and conducted the phytochemical test. DM and SLPK drafted the article. SLW and AK corrected the final manuscript. All authors approved the final version of the manuscript.

Conflict of interests
The authors declare no conflict of interest.

Ethics considerations
Prior authorization for the use of laboratory animals was obtained from the Cameroon National Ethics Committee (Ref. N°. FWIRB 00001954).

Funding/Support
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References